International Journal of Ayurvedic and Herbal Medicine 11:6 (2021) 4034–4068

Journal homepage: <u>http://interscience.org.uk</u> DOI:10.47191/ijahm/v11i6.02 Impact Factor: 6.012

A Comparative Review on Medicinal Plants Used for the Liver Disorders as in Ayurved, Siddha and Unani [ASU] Systems of Medicine

Part 2 - Phytochemical and Pharmacological Aspects

Dr. Bhatt Narendra¹, Dr. Deshpande Manasi²

¹Consultant- Ayurveda, Research & Industry, CRIA Consultants Pvt. Ltd. 15, Bachubai Building, J.Bhatankar Marg, Parel, Mumbai 400 012 ²Professor and Head, Department of DravyagunaVigynan, Bharati Vidyapeeth [Deemed to be University], College of Ayurved, Pune, 411043

ABSTRACT: Liver disorders mostly represent a prolonged dysfunction associated with different routes of cellular and biological alterations due to variety of reasons. Present treatments are found to be of limited use. Indigenous systems of medicine Ayurved, Siddha and Unani are extensively used to provide relief in variety of liver diseases. The understanding of liver functions, its dysfunctions and treatments offered have been of research interests. A series of review papers have been published by the authors covering different aspects including classical, proprietary and This review paper- part II is the outcome of the systematic effort and analyses of hepatoprotective formulations and ingredients with the updated information on phytochemical and pharmacological- safety aspects. This strongly emphasizes on the role of ASU products and plants exhibit huge translational potentials to evolve new path to understand and develop new safe and effective therapeutic approaches and drugs to treat liver disorders.

KEY WORDS: Liver disorders, Ayurved, Siddha, Unani, Medicinal plants, pharmacology

Patented products. A summarized outcome on 25 potential medicinal plants out of 106 has been published as part I.

BACKGROUND

Worldwide the liver diseases are significant cause of morbidity and mortality. Multiple etiological factors are cause of liver dysfunction that can lead to complex conditions although the rates of progression and clinical course may be different^[1].

Ayurved, Siddha and *Unani* [ASU] are indigenous systems of medicine of India that provide descriptions of different types of liver diseases and offer therapeutic approaches, products, and ingredients of natural origin to treat them. Hundreds of liver products based on these systems are prescribed by physicians and used as remedies by people.

A series of reviews undertaken by the *Bhatt et al* provide major information about hepatoprotective herbal drugs used in ASU traditional medicine for the treatment of liver diseases ^{[2-7].}

METHODOLOGY

In a recent review, the ccontextual and clinical aspects of of 106 plants used singly or in combinations were scrutinized to prepare a priority list of 25 plants from ASU systems for treatment of liver disorders ^[8-19].

This review is based on phytoconstituents and preclinical data of medicinal plants used in liver disorders obtained through updated Google and PubMed searches to help to select high potential ingredients or their combinations.

LIVER AND LIVER DISORDERS IN ASU SYSTEMS]^[20-24]

The significance of the liver in the context of blood-fluid, which embodies one of the main humoral or functional systems of human biology, explicitly explained in the ASU systems. Any functional disturbance of the humoural system or the temperament may develop into various types of liver disorders and on the other hand, any impairment of liver function can affect one or more of the body functions. These pathological conditions vary in disposition and are a result due to weakened hepatic activity, dullness, inflammation, obstruction, trauma and such others.

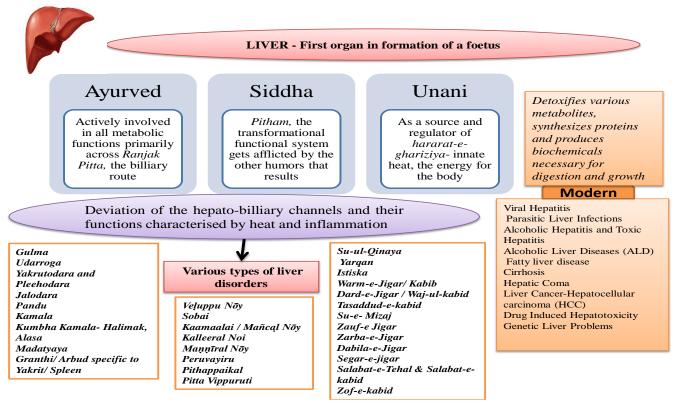


Fig 1- Summarize the Liver concept and liver diseases in ASU system of medicine

HIGH POTENTIAL MEDICINAL PLANTS

The potential **twenty-five** plants were screened and scrutinized based on published literature for scientific assessment of phytoconstituents and pre-clinical activities related hepatic functions listed in figure 2.

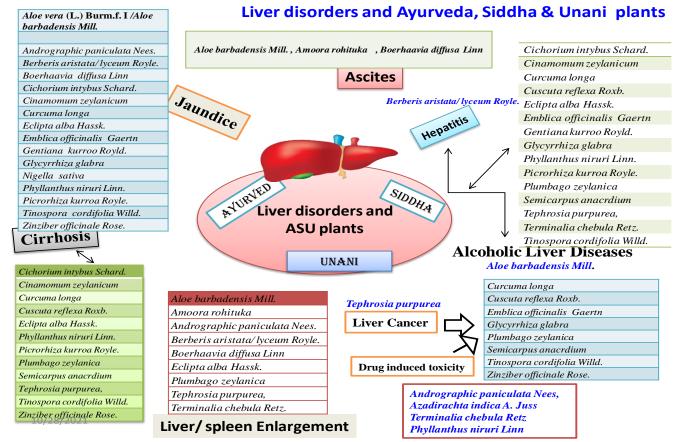


Figure 2: High Potential Medicinal Plants from ASU Systems for Liver Disorders

RESEARCH SIGNIFICANCE OF PHARMACOLOGICAL- PRECLINICAL AND SAFETY STUDIES

Aloe vera (L.) Burm.f. I /Aloe barbadensis Mill (Liliaceae)

Aloe, amongst one of the most well-known home remedy plant is used in ASU traditional systems of medicine for improving liver function by stimulating release of the bile. It contains Phenolic, Glycosides, Anthrones, Barbaloin, Aloe-emodin 12 anthraquinones.

Aloe *juice* has ameliorative effect in malathion induced hepatotoxicity in rabbits ^[25]. Hepatoprotective effect of the plant (stem) exhibited in *ethanol* and *aqueous extracts* in paracetamol induced liver damage ^[26], while petroleum ether, chloroform and methanol extracts in CCL₄ induced hepatotoxicity ^[27]. *Polysaccharides AVGP* in chronic alcohol feeding mice regulated hepatic expression of lipolytic genes (AMPK- α 2 and PPAR- α), accelerated lipolysis and inhibited inflammatory response ^[28].

Whole leaf extract showed carcinogenic activity in rats, as a possible human carcinogen (Group 2B); a dosedependent decrease in the viability in HeLa and HepG2 cells. Cytotoxicity and genotoxicity study of *whole extract* showed a positive response at lower concentrations than the decolorized extract in the mouse lymphoma assay (MLA). Reproductive toxicity - sperm damage, haematological changes, inflammation, mortality was observed after a chronic oral ingestion of 100 mg/kg *extract* per day, for 3 months^[29].

Amoora rohituka (Roxb.) Wight&Arn./Tecoma undulata (Sm.) G.Don (Bignoniaceae)

Bark of *Rohitak* having blood purifying and cholagogue properties. It contains tecomin (veratroyl β -D-glucoside), n-triacontane, n-heptacosane, n-nonacosane, n-triacontanol, n-octacosanol; β -sitosterol. Its hepatoprotective activity observed in the *ethanolic extract of bark* in paracetamol induced hepatic damage^[30], *methanol extract* of leaves in alcohol-induced hepatotoxicity in rats^[31]. It prevented changes in metabolic

enzyme concentrations as well as diminished the destruction of liver cell architecture initiated by administration of $CC14^{[32]}$.

Andrographis paniculata Nees (Acanthaceae)

The king of bitters has strong liver stimulating activity to expel out excess bile, *pitta* from the body. It helps regulating hepatic function by reducing inflammatory biological factors. 55 *ent*-labdane diterpenoids, 30 flavonoids, 8 quinic acids, 4 xanthones, and 5 rare noriridoids are present in the plants.

The *aqueous extract* reduces CCl₄, paracetamol induced hepatotoxicity ^[33], prevents liver damage ^[34], antioxidant effect ^[35]. *Intra-peritoneal* in the albino rats the *extract* has choleretic effect with qualitative changes and increase in the bile secretion ^[36]. In galactosamine and paracetamol intoxication, it helps normalize the biochemical parameters ^[37-38], hepatic microsomal drug metabolizing enzymes ^[39-40]. The diterpenes *andrographolide* (I), *andrographiside* (II) and *neoandrographolide* (III) exhibited protective effects on CCl₄ or tert-butylhydroperoxide intoxication hepatotoxicity in mice ^[41].

<u>No acute or sub acute toxicity</u> was observed in LD - 50 studies with andrographolide and derivatives ^[42].

Azadirachta indica, A. Juss (Meliaceae)

The world famous Margosa tree for its antipollution attributes amongst many known properties is recognized for its anti-parasitic, healing, and hepato-protective effects where its anti-inflammatory activity regulates impaired levels of liver enzymes. The most important active constituents are azadirachtin, nimbolinin, nimbin, nimbidin, nimbidol, sodium nimbinate, gedunin, salannin, and quercetin.

Healing potential of the powder of Neem *leaves* on liver parenchyma with regeneration of liver cells has been studied in CCl₄ induced hepatotoxicity ^[44]. *Aqueous leaf extract* is useful in prevention and reversal of the hepatotoxic damage due to antitubercular drugs ^[45] and has shown hepatoprotective activity in paracetamol induced hepato-toxicity in albino rats ^[46-47].

Ethanolic, methanolic, aqueous leaf extracts have shown hepatoprotective effects in CCl₄ induced hepatotoxicity ^[48-49] and the *methanolic extract* alleviates cisplatin-induced damage and oxidative stress in liver ^[50].

Azadirachtin-A, *Nimbolide* reduced hepatocellular necrosis in CCl₄ induced hepatotoxicity and has antioxidant effect^{[51].} Oral administration of *water extracts of leaves and seeds* exhibited <u>dose related toxicity</u>-with decrease in death percentage from 100 % to 33 % in rats and rabbits; <u>variable solvents and methods of preparation could affect the levels of toxicity</u>^{[52].}

Nimbolide and *nimbic acid* are *toxic* to mice only when given i.p. and i.v. but they are less toxic to rats and hamsters by the oral route. It observed dose-related <u>pharmacotoxicity</u> symptoms [possible dysfunctions in kidney (tubular necrosis), small intestine (hemorrhagic necrosis), pancreas (acinar cell necrosis) and liver (mild fatty infiltration and focal necrosis^[53]. Neem *oil* which is regularly used <u>did not have any toxicity</u> in 90-day sub chronic toxicity studies as in serum biochemistry, organ weight and histopathology^[54-55].

Berberis aristate DC. (Berberidaceae)

A strong astringent Berberis plant used effectively in treating anorexia and dysentery is a hepato-stimulant having cholagogue effect. It exhibits suppressing action on drug metabolizing enzymes and improves the functional recovery of beta cells. The plant contains barberine, oxyberberine, berbamine, aromoline, karachine, palmatine, oxyacanthine and taxilamine.

The hepatoprotective effects of its *crude powder* and *methanolic extract* are observed in paracetamol induced hepato-toxicity^[56], the *fruit extract* in paracetamol-induced liver damage^[57].

Berberine hydrochloride in Hepg2 Cancer cell exhibited regulation of prostaglandin E2 (PGE2) signalling pathway line via PGE2 and Cox2 inhibition^[58].

Berberine showed antioxidant-hepatoprotective activity in CCl₄ induced hepatotoxicity ^[59], and in HFD - high fatty diet - induced rat model where methylation of the MTTP - microsomal triglyceride transfer protein - helped alleviate fatty liver^[60].

Powdered root, extract and *pure berberine* showed *sub-acute toxicity* on rat, rabbit and mice. The sub-acute concentrations of *berberine* lead to <u>altered liver function</u>, <u>gastric troubles</u>, <u>hepato-hemato-toxicity</u>, <u>hemorrhagic inflammatory consequences</u>, <u>damage to immune cells and induced apoptosis</u>. The in vivo and in vitro studies have reported that *sanguinarine* may induce <u>apoptosis and adversely influence the embryonic</u> <u>development</u> (both in the pre-implantation and post-implantation conditions) of mouse ^[61].

Boerhavia diffusa, Linn.(Nyctaginaceae)

Punarnava having large quantity of *nitrate content* has strong diuretic and anti-inflammatory activity and is frequently used in cirrhosis and ascites with advanced structural changes to reduce the functional liver burden. It contains b-Sitosterol, a-2-sitosterol, palmitic acid, tetracosanoic, hexacosonoic, stearic, arachidic acid, urosilic acid, Hentriacontane, b-Ecdysone, triacontanol etc.

Aqueous extract has shown antioxidant and hepatoprotective effects in acetaminophen, CCl₄^[62], paracetamol ^[63] and ethanol induced hepatotoxicity ^[64-65]. It has shown increase in lipid peroxides further increasing in activities of the superoxide dismutase, catalase, glutathione peroxidase and glutathione-S-transferase and reduced glutathione levels ^[67]. Its *hydroalcoholic extract* showed hepatoprotective activity in d-galactosamine induced hepatotoxicity ^[68].

Aqueous extract of leaves in <u>sub-chronic toxicity studies</u> on liver enzymes and haematological parameters was observed to be <u>non-toxic^[69]</u> and in <u>teratogenic studies no foetal anomaly</u> were detected with its *ethanol extract* ^[70].

Cichorium intybus L. (Compositae)

Cichori plant is traditionally used in the treatment of almost all kinds of liver disorders like sluggish liver, enlargement of spleen, billiary stasis (stoppage of bile) and jaundice. It's cooling effect increases secretion of bile by reducing inflammation, remove obstruction, and helps in stomach pain and obesity. Inulin, coumarins, tannins, monomeric flavonoids, sesquiterpene lactones are some of the major phyto-compounds.

Hepatoprotective effects of *aqueous and alcoholic extracts* were observed in acetaminophen and CCL₄ induced liver damage in mice with decrease in the levels of increased hepatic enzymes ^[71-76]. Similarly anti-hepatotoxic effects with decrease in the hepatic enzyme levels were found with the *root and root callus extracts* in CCL₄ induced liver damage in rats ^[77].

Cichotyboside and its *ethanol extract* also improved liver enzyme levels in CCl₄ induced hepatotoxicity^[78-79]. Phenolic acid-rich *seed extract* helped in restoration of normal levels of corresponding proteins in oleic acid-induced non-alcoholic fatty liver disease (NAFLD)/steatohepatitis (NASH) through modulation of PPAR-alpha and SREBP-1hepatic steatosis in vitro and in vivo –to up-regulate the expression of SREBP-1c and PPAR- α genes^[80].

There is a slight potential for <u>sensitization via skin</u> contact with the drug^[81].

Cinnamomum zeylanicum, Blume (Lauraceae)

Essential oils of cinnamon bark and leaf are widely used in food flavours, cosmetics and pharmaceuticals. E)-Cinnamaldehyde is the main component of cinnamon bark oil and eugenol the main component of the leaf oil. Cinnamon bark extract and Oil Cinnamaldehyde have shown antioxidant and hepatoprotective activity in CCL₄^[82] and paracetamol induced hepatotoxicity^[83].

Cinnamon and its main constituents <u>ameliorate the toxicity</u> in liver, kidney, blood, brain, embryo, reproductive system, heart, spleen through antioxidant effects, modulation of CK-MB, LDH, TNF- α , IL-6, mitogen-activated protein kinase (MAPK), and nuclear factor- κ B (NF- κ B) signalling pathways in vitro and animal studies on the protective effects against natural and chemical toxins^[84].

Curcuma longa, L. (Zingiberaceae)

Golden spice of India has been used as a household remedy for various diseases, including biliary disorders. Turmeric contains curcuminoids (mixture of curcumin, demethoxycurcumin and bisdemethoxycurcumin). *Aqueous extract* of Turmeric has shown antioxidant-hepatoprotective activity in bleomycin-induced & ethanol-induced hepatotoxicity^[85-86], *ethanol extract* in paracetamol induced toxicity^[87].

Curcumin has shown variety of hepatoprotective in CCL₄ and lead induced hepatotoxicity^[88], antioxidant activity^[89], and its anti-inflammatory effect helped prevent the progression of irreversible liver dysfunction in dimethylnitrosamine (DMN) induced liver cirrhosis in rat model^[87,90]. *Curcumin* exhibited enhanced hepatoprotective effects inhigh fat diet (HFD) NAFLD rat model ^[91] and billiary duct ligation (BDL) and also in marine model of non-alcoholic steatohepatitis methionine and choline deficient diet [MCD] with prevention of accelerating of oxidative associated liver diseases^[92-95].

No adverse effect level (*NOAEL*) was observed for <u>reproductive toxicity</u> of *curcumin*, fed in the diet for two successive generations^[96-97].

Curcumin in wistar rats <u>did not show</u> significant effect on the incidence of micro nucleated polychromatic erythrocytes, structural and numerical aberrations in bone-marrow chromosomes, pregnancy rate, and number of live and dead embryos, total implants and mutagenic index. <u>No toxic effects</u> were stated at doses of 3.5 g/kg given for 3 months in rats, dogs, and monkeys. The use of curcumin in humans by oral administration observed that 1.5 g of turmeric powder per day (about 150 mg of curcumin, average consumption in India) <u>did not exhibit any side effects</u> in humans ^[98].

Synthetic Curcumin did not show any mortality or 'No adverse effect level[*NOAEL*]', <u>toxic effect or</u> <u>genotoxicity</u> in 90-day repeated-dose at daily doses of 250, 500, or 1000 mg/kg body weight/day, administered by gavages in a split dose and repeated-dose^[99].

Cuscuta reflexa Roxb. (Cuscutaceae)

C. reflexa contains cuscutin, amarbelin, β -sitosterol, stigmasterol, kaempferol, dulcitol, myricetin, quercetin, coumarin and oleanolic acid and has anti-tumor, anti-oxidant and anti-inflammatory activities.

Alcoholic extract of stem, whole plant showed hepatoprotective activities in paracetamol, carbon tetrachloride, ethanol induced hepatotoxic rat models^[100-101]. The hepatoprotective effects were observed in *chloroform and ethanol ethanolic extract* in paracetamol and *alcoholic and aqueous extracts* of *stem* in thioacetamide induced liver damage in rats^[102-103].

Eclipta alba f. prostrata (L.) Hassk.(Asteraceae)

Eclipta leaves are most commonly used in variety of liver disorders in all the three systems. A broad range of chemical components including alkaloids, alkenynes, cardiac glycosides, steroids, triterpenes, phytosterol, flavonoids, coumestans, glycosides, triterpinoids, saponins have been extracted from the plant.

Fresh powder, aqueous, hydroalcoholic and alcoholic extracts of leaves have repetitively shown hepatoprotective effects in CCL₄^[104-108], alcohol^[109] and paracetamol^[110-112], induced acute or chronic liver damage in rats and anti-hepatotoxic activity in various hepatotoxins, aflatoxin induced acute hepatitis^[113-115] by restoration of Na+K+ATpase activity and regulating hepatic microsomal drug metabolising enzymes. Also, *methanol extract of leaves* and *chloroform extract of roots* normalized serum GOT, GPT, ALP, and bilirubin levels in CCL₄ induced hepatotoxicity^[116].

Coumestans (wedelolactone and dimethyl wedelolactone exhibited anti-hepatotoxic stimulatory effect on liver cell regeneration in assays employing CCl₄, GalN and phalloidin-cytotoxicity in rats ^[117]. *Wedelolactone, luteolin,* and *apigenin* also exhibited dose-dependent inhibition of HCV inhibitory activity by

replication in vitro and anti-HCV (hepatitis C virus) in the cell culture system^[118].

Emblica officinalis Gaertn

Emblica officinalis Gaertn or *Phyllanthus emblica Linn*, Indian gooseberry is one of the most important medicine, dietary and one of the ingredients of '*Triphala*' (combination of three fruits) that is most commonly used for a wide range of cathartic effects.

Amla fruit contains higher quantity of polyphenols like gallic acid, ellagic acid, different tannins, minerals, vitamins, amino acids, fixed oils, rutin and quercetin.

Scientific studies have shown that *Amalaki* is effective in preventing/ameliorating the toxic effects of hepatotoxic agents like ethanol, paracetamol, carbon tetrachloride, heavy metals, ochratoxins, hexachlorocyclohexane, antitubercular drugs and hepatotoxicity resulting from iron overload. It has beneficial effects on liver function, hyperlipidemia and metabolic syndrome^[119].

Fruit extract has range of antioxidant and hepatoprotective effects as observed in alcohol induced hepatotoxicity in rats^[120-121], arsenic induced hepatopathy in adult Swiss albino mice^[122], chronic and preventive, pre-fibrogenesis of liver cells, with CCl₄ and thioacetamide induced liver damage^[123-125]. Further *pre-treatment with seven consecutive* days inhibited hepatotoxicity in CCl₄ induced toxicity in wistar rats ^[126]. 50% *hydro-alcoholic extract* of the fruit expressed stabilizing, antioxidative and cytochrome (CYP) 2E1 inhibitory effects in anti-tubercular drugs-induced hepatotoxicity ^[127]. In paracetamol and CCl₄ induced hepatotoxicity the *tannins, flavonoids, terpenoids, alkaloids of fruits* showed offset of necrosis with appearance of normal hepatocytes, and consequent appearance of leucocytes^[128-129].

Amla supplementation counteracts NDEA induced liver injury via its antioxidant, anti-inflammation, antiapoptosis, and anti-autophagy properties ^[130]. Its extract significantly inhibited hepato-carcinogenecity induced by NDEA in a dose dependent manner ^[131].

Glycyrrhiza glabra L. (Fabaceae)

World over the liquorice is a known home remedy including liver ailments like non- alcoholic fatty liver. Its extract reduces the concentration of ALT and AST, indicating its beneficial effect on the liver functions. Liquorice roots contain more than 20 triterpenoids and nearly 300 flavonoids. Glycyrrhizin and glycyrrhetinic acid are the main components.

Aqueous, hydro-methanolic and ethanol extracts of liquorice have variably shown hepatoprotective effects in CCl₄-induced hepatotoxicity, oxidative and in-vitro hepatocyte damage in rats^[132-135] and ameliorating liver functions as well as restoring hepatic tissue in acute liver diseases^[136]. *Methanol extract of roots* exhibited

glucuronidation in rat metabolism of acetaminophen in male Sprague-Dawley rats with increased the cumulative billiary and urinary excretions of acetaminophen, glucuronide conjugate ^[137].

Glycyrrhizin-Secretion of HBsAg and accumulated it dose-dependently in PLC/PRF/5 cells, chronic hepatitis B- suppressed the intracellular transport of HBsAg at the trans-Golgi area after O-linked glycosylation and before its sialylation, improved the liver histology, intracellular transport and suppresses hepatitis B virus surface antigen, prevents development of hepatocellular carcinoma in chronic hepatitis^[138-139].

 8β -glycyrrhetinic acid, an aglycone of glycyrrhizin decreases the expression of P450 E1 in CCl₄ induced liver injury thereby protecting the liver ^[140].

Glycyrrhizic acid in Concanavalin A -induced mouse model had effect on mice CD4(+)T cells in livers and spleens that showed inhibition of hepatic fibrosis^[141].

Compound *glycyrrhizin* tablet improved the liver dysfunction that augmented the entire cytotoxic function mediated by hepatic lymphocytes inhibiting the cascade in HIV induced apoptosis. It inhibited cross-linking between HIV spikes and CD₄, a cell surface molecule, by suppressing CD₄ expression, and suppresses apoptosis ^[142]. *Glycyrrhizin* and *glycyrrhetinic acid* with interferon induction in mice showed activation of macrophages and augmentation of NK activity through the action of the induced IFN^[143].

Prolonged use of excessive doses leads to pseudoaldosesteronism indicated with potassium depletion, sodium retention, oedema, hypertension and weight gain^[144].

Luffa echinata Roxb. (Cucurbitaceae)

Various parts of Luffa are useful for the treatment jaundice, enlargement of liver and spleen, cirrhosis, dropsy. The active constituents are cucurbitacin, saponin, echinatin, β -Sitosterol, oleanolic acid and flavonoids.

Petroleum ether, acetone, methanolic and ether extracts of Luffa showed hepatoprotective activity in CCl₄ induced hepatotoxicity in albino rats ^[145-146]. Its crude extract has shown significant improvement in biochemical parameters in liver injuries ^[147]whereas the aqueous extract significantly lowered serum bilirubin levels in chlorpromazine induced jaundice in rats^[148]. Its Hydro-alcoholic extract showed anti-hyperglycaemic activity along with improvement in renal and hepatic functions ^[149].

Moringa oleifera Lam (Moringaceae)

Drum stick plant is widely used as nutritional herb and contains rich source of the vitamin A, vitamin C and milk protein. Different types of active phytoconstituents like alkaloids, protein, quinine, saponins, flavonoids, tannin, steroids, glycosides, fixed oil and fats.

Extract showed protection against cadmium-induced toxicity in rats ^[150]. Aqueous extract restored the lead perturbations through reduction of oxidative stress—induced DNA damage via amelioration of NF-kB and TNF- α which kept hepatocyte integrity and reduced serum hepatic enzyme activities ^[151]. Ethanolic extract of leaves enhanced the recovery from hepatic damage induced by antitubercular drugs ^[152], prevented and improved liver damage^[153]. Methanolic leaf extract showed antioxidant and dose-dependent hepatoprotective activities in CCl₄ model ^[154].

Nigella sativa L. (Ranunculaceae)

Bleck seed has been used as antihypertensive, liver tonics, diuretics, digestive, anti-diarrheal, appetite stimulant, analgesics, anti-bacterial. The active compounds are thymoquinone, thymohydroquinone, dithymoquinone, p-cymene, carvacrol, 4-terpineol, t-anethol, sesquiterpene longifolene α -pinene and thymol *etc*

Basal diet with *Nigella sativa* addition in lead nitrate induced toxicity showed protective effect against toxic effect of lead on liver and kidney tissues ^[155]. Aqueous extract showed hepatoprotective effects in nimesulide and paracetamol induced hepatotoxicity ^[156-157]. Volatile oil in CCL₄ induced hepatotoxicity decreased the lipid per-oxidation and liver enzymes, increased the anti-oxidant defence system activity ^[158].

Hydro-alcoholic extract 0.2 mL/kg by intraperitoneally administration in the rats protected the rat liver against to hepatic ischemia-reperfusion injury ^[159].

Hepato protective activities of water extract were observed in rats with CCl₄-challenged damage^[160], with 5-(Aziridin-1-yl)-2,4-dinitrobenzamide (CB 1954) hepatotoxicity^[161], and Balb/C mice transplanted with the 66cl-4-GFP cell line for antitumor activity^[162].

Thymoquinone from NS showed hepato protective effect in acetaminophen induced hepatotoxicity ^[163-164]& supplementation prevents the development of DENA-induced initiation of hepatic carcinogenesis by decreasing oxidative stress and decreased mRNA expression of GSHPx, CAT and GST ^[165].

Thymoquinone orally as well as intraperitoneal, LD_{50} of both in mice and rats by the method of Miller and Tainter, was found relatively a safe compound, particularly when given orally^[166-167]. Fixed oil of NS did not show any - LD_{50} acute or chronic toxicity^[168].

Phyllanthus niruri Linn& species (Euphorbiaceae)

Phyllanthus is probably the most widely explored genus for liver disorders. Recognized for its role in viral hepatitis, *P. niruri* is used in ASU systems for different biliary conditions and in preventing gallbladder calculi. It contains flavonoids, terpenoids, alkaloids, polyphenols, lignans, tannins, saponins, coumarins.

Hot water extracts showed antioxidant activity and the protective effect in paracetamol-induced liver damage ^[169] [240]. Aqueous leaves extract modulated the expression of matrix metalloproteinases in alcohol and thermally oxidized polyunsaturated fatty acid-induced hepatic fibrosis ^[170].

In CCl₄ induced liver damage fresh leaves juice, aqueous and methanolic extracts of seeds showed hepatoprotective effects^[171-172] and ethanolic extract of aerial parts showed subsequent recovery towards normalization in Swiss strain female albino mice^[173-174].

Hepatoprotective effects were also observed in ethanolic extract of whole plant except root in aflatoxin B_1 -induced liver damage in mice ^[175] and in thioacetamide induced liver cirrhosis in rats ^[176]. Hepatoprotective effects of methanolic, aqueous extract of the leaves and phenolic constituents were observed in ethanol-induced oxidative damage in adult male wistar albino rats with glucuronidation in rats ^[177]. Extracts significantly inhibited hepatocarcinogenecity induced by N- nitrosodiethylamine in a dose dependent manner ^[131].

Hepatoprotective and mechanistic benefits of Corilagin C₁, Isocorilagin C₂, Brevifolin C₃, Quercetin C₄, Kaempferolrhamnoside C₅, Gallic acid C₆, Brevifolin carboxylic acid C₇-CCl₄ induced toxicity in Clone-9 and Hepg2 cell lines were observed ^[178].

Protein isolate had hepatoprotective effect in acetaminophen and CCL₄-induced toxicity^[179-180].

Phyllanthin showed antioxidant capability of rat hepatocytes including level of total glutathione, and activities of SOD and glutathione reductase ethanol-induced hepatotoxicity in rats^[181].

Picrorhiza kurroa Royleex Benth. (Scrophulariaceae)

Rhizomes of *Kutaki* are bitter in taste, cooling and use for removal of excessive *Pitta* from the body via colon. It helps in restoration of Liver functions by overcoming fatty liver changes.

The bioactive compounds of plants are iridoids-icroside I, picroside II, picroside III, picroside IV, kutkoside, pikuroside, cucurbitacin and acetophenones.

Aqueous-ethanolic extracts protected hepatotoxicity induced by acetaminophen in Cockerels^[182],aqueous extract in liver slice culture system; ethanol restored the activities of antioxidant enzymes and significantly reduced lipid peroxidation ^[183]. Its extract significantly inhibited hepatocarcinogenecity induced by N-nitrosodiethylamine in a dose dependent manner^[131].

Hydroalcoholic extract in non-alcoholic fatty liver disease showed reversal of the fatty infiltration of the liver and lowering of the quantity of hepatic lipids ^[184].

Picoliv has shown significant hepatoprotective effects in alcohol and paracetamol induced hepatotoxicity in rats ^[185-187], aflatoxin B₁-induced liver damage in mice ^[188] and choleratic effect with increase in bile flow & change in the physical properties of the bile secretion ^[189]. *Picroliv or Kutkin* again showed hepato-protective and immune-modulator activities in D- galactosamine, paracetamol, thioacetamide and CCl₄ induced hepatic damage ^[190].

In hepatic amoebiasis associated with CCl₄ toxicity it showed hepato-generative effect^[191] and decreased the levels of lipid peroxides and hydroperoxides and facilitated the recovery of superoxide dismutase and glycogen^[192-194].

The hepatoprotective and bile flow enhancing activity of Kutkin- iridoid glycoside have been demonstrated^[195].

Piper longum L.(Piperaceae)

Long pepper (one of the three ingredients of *Trikatu-* pungent herbs), used as a *Rasayana*, is useful in respiratory & digestive disorders. The fruits contain volatile oil, resin and alkaloids piperine and piperlonguminine.

Traditional milk extract– dried fruits boiled with milk – showed protective effect in CCl₄ induced hepatotoxicity ^[196]. *Piperine-* major active constituent-bio enhancer activity- protects against hepatocellular injury and fibrosis ^[197-198].

Piperine showed showed regeneration process by restricting fibrosis in CCl₄ and tertiary butylhydroperoxide induced hepatotoxicity; but offered no protection against acute damage or against cirrhotic changes ^[199].

 LD_{50} values of Piperine on adult male mice showed that most animals died of respiratory paralysis within 3– 17 min, while in sub-acute toxicity studies, the rats died within 1–3 days after treatment. In histopathology severe haemorrhagic necrosis and oedema in gastrointestinal tract, urinary bladder and adrenal glands observed. Death may be attributable to multiple dysfunctions in organs^[200].

Plumbago zeylanica L.(Plumbaginaceae)

Root of Plumbago, is considered a strong digestive stimulant that helps relieve indigestion, gas, bloating, cramping and constipation, a detoxifier - blood purifier being effective against liver flukes. Roots of the plant contain an acrid crystalline principle called plumbagin, chloroplumbagin and biplumbagin.

Hepatoprotective effects are observed of rhizome extracts in CCl₄-induced ^[201-202]] and Petroleum ether extract in paracetamol induced liver damage ^[203].

In acute and sub-acute-toxicity studies with petroleum ether, acetone, and the hydro alcoholic extracts no mortality was observed with acute safe dose but liver and kidney were adversely affected following the sub-acute administration of root extract in rats^[204].

Swertia chirata Buch Ham(Gentianaceae)

It is a bitter tonic, carminative, laxative, anti-pyretic, febrifuge, anti-periodic, anti-inflammatory, stomachic, and anthelmintic used in treating various types of fevers, liver diseases, piles, skin diseases, ulcers, and

diabetes. The main chemical ingredients are Swertiamarin, Amarogentin, Swechirin, Mangiferin, Sweroside, Gentianine, Amaroswerin, Oleanolic acid, Swertanoone, Ursolic acid.

Ethanolic extracts improvised levels of serum marker enzymes in paracetamol-induced hepatotoxicity^[205-206]. *Syringaresinol-* Methanolic extract showed anti-hepatotoxic activity against CCL4, galactosamine and paracetamol induced liver toxicity^[207-208].

Tephrosia purpurea (L.) Pers.(Fabaceae)

It is cholagogue, deobstruent, diuretic, tonic and laxative; and used in enlargement and obstruction of liver. It contains rutin, purpurin, purpurenone, quercetin, retinoids, deguelin, elliptone, rotenone, sitosterol, and tephrosin. It protects the liver against drug induced oxidative damage probably by increasing antioxidative defense activities.

Variety of hepatoprotective effects of whole plant aqueous extract in paracetamol-induced^[209], Ethanol extract of roots in CCl₄ induced^[210], Hydro-alcoholic extract against sodium arsenate induced sub-acute toxicity^[211] and Aqueous-ethanolic extract of aerial parts in thioacetamide induced liver cirrhosis in rats^[212] have been observed.

Terminalia chebula Retz (Combretaceae)

Haritaki is principal ingredient of *Triphala* (combination of three fruits) that is recognized for a wide range of cathartic effects. It improves metabolism and provides beneficial effects to colon, liver, spleen and lungs.

T. chebula is of pyrogallol (hydrolysable) type; contain 14 components of hydrolysable tannins. Water extract prevented liver toxicity in anti-tuberculosis drug-induced toxicity (Sub-chronic administration of rifampicin, isoniazid and pyrazinamide in combination)^[213] and in den induced hepatocellular carcinogenesis in experimental rats^[214].

It prevents acute and severe liver injury, inhibition of oxidative stress and inflammatory cytokines with membrane stabilizing activities in C57/BL6 mice model of tert-butylhydroperoxide induced acute liver injury ^[215]. It exhibited chemo preventive potential by estimating the levels of lipid peroxidation and assaying activities of various marker enzymes in paracetamol-induced liver damage ^[216].

Ethanolic fruit extract showed hepatoprotective effect in ethanol-induced hepatotoxicity in rats ^[217] whereas *chebulic acid* showed antioxidant effect in isolated rat hepatocytes ^[218], tannins flavonoids expressed protective mechanism in N-nitrosodiethylamine induced-hepatocellular carcinoma ^[219].

Crude alcoholic extract did not exhibit any *cytotoxic effect* to fresh sheep erythrocytes cytotoxins in alliums model or any *genotoxic effect* in either vitotox test or Ames assay^[220-221].

Tinospora cordifolia (Willd.) Miers (Menispermaceae)

Guduchi is probably known as one of the best-known immunomodulatory plant from Ayurveda. A variety of active components like alkaloids, steroids, diterpenoid lactones, aliphatics, and glycosides have been isolated from different parts of the plant.

Whole plant aqueous extract benefitted in bile duct ligation induced jaundice in rats ^[222]. Hepatoprotective effect of water extract was observed in CCL₄ induced liver toxicity in goat ^[223]. Aqueous extract showed protective effects in hepatic and gastrointestinal toxicity induced by chronic and moderate alcoholism ^[224]; significant improvement in Kupffer cell function using carbon clearance test showed a trend towards normalization ^[225]. Aqueous, petroleum ether and ethanol extracts of various parts of the plant prevented liver toxicity in CCL₄ damage ^[226-227]. Ethanolic extract showed hepatoprotective effect in CCl₄ induced liver damage in mice, rat, and rabbit ^[228-231]. Protective role of stem and leaves extract was observed in lead nitrate

induced toxicity^[232], aflatoxin-induced nephrotoxicity on thiobarbituric acid reactive substances levels with increase the level of GSH, ascorbic acid, protein, the activities of anti-oxidant enzymes^[233].

Zingiber officinale Roscoe (Zingiberaceae)

Ginger is a spice consumed worldwide for culinary and medicinal purposes contains Phenols, Oleoresins, Zingiberene, Zingib

Hepatoprotective and antioxidant effects of ethanolic extract of Ginger rhizomes have been observed in CCl₄, acetaminophen, paracetamol and thioacetamide induced liver toxicities in rats^[234-237].

Dry ginger with essential oils – phenols, flavonoids components studied in DEN - induced hepatocellular carcinogenesis in rats reduced the severity cytotoxicity and anticancer potential against HepG2 cell line^[238]. The detoxicating action of Ginger in liver was found to be more effective therapeutically on cadmium toxicity, accumulated cadmium, than prophylactically^[239]

Significance of pharmacological models and plants studied

Animal models have played an important role in biomedical research as crucial tool for study and understanding the pathogenesis of several liver diseases, help to develop new pharmacologic treatments. Each model in vivo and in vitro has several characteristics, advantages and limitations.

Table 1. summaries of several experimental models for hepatoprotective and hepatotoxicity activities and plants listed from above observations

No.	Model & Plants	Relevance	Possible activities	Limitations
1.	CCL ₄ ^[240]	Metabolized by	Reactive oxidative free	Prolonged treatment
	(21)	cytochrome P-450 in	radical-initiates lipid	promotes severe
		endoplasmic reticulum,	peroxidation	cirrhotic changes with
		mitochondria with the		significant development
		formation of ccl3o		of Ascites
2.	Paracetamol	Analgesic- antipyretic	Produces cell necrosis	Produces Acute liver
	[240]	lipid peroxidative		damage in high dose
	(18)	degradation of		
		glutathione level		
3.	Acetaminophen	Encountered drug	Reactive metabolite	Overdose leads to liver
	[241]	hepatotoxicity	formation, Mitochondrial	failure, death
	(6)		damage, DNA	
			fragmentation, Necrosis	
4.	Ethanol ^{$[242](5)$}	Steatosis, elevation of	Mild elevation of serum	Lack of ability to
		CYP2E1, oxidative	ALT, AST, Low levels of	develop ALD
		stress, increases gut	liver inflammation	
		permeability, activates		
		LPS-TLR4-Kupffer cells		
5.	$Alcohol^{[243]}(5)$	Changes in membrane	Mild liver injury, steatosis,	Dehydration, low blood
		lipid composition,	low-grade liver	alcohol level,
		fluidity, increase hepatic	inflammation	inadequate nutrition
		lipid peroxidation		

6	Thioscotom: do [24	Interfore the movement	Mombrono iniumy raduces	Long time to develor
6.	Thioacetamide ^{[24} 4] (5)	Interfere the movement of RNA from the nucleus to cytoplasm	the number of viable hepatocytes, rate of oxygen consumption, decreases the volume of bile	Slow reversibility
7.	Anti-tuberculosis drug- Isoniazid and rifampicin ^[245] (4)	Oxidative stress, Produced steatosis, increased SDh, indicated mitochondrial injury	Adverse effects on the liver, ranging from mild transient elevations in aminotransferases to overt hepatitis	Pathogenesis underlying hepatotoxicity poorly understood
8.	Aflatoxin B ₁ / Hepatotoxin ^[246] (4)	Toxin-dependent liver injury, hepatic GH- resistance	Dose-dependent wasting, stunting, liver pathology, suppression of hepatic targets of GH signalling	Epoxide from binds with protein may lead to Compensatory liver hyperplasia, promote the incorporation of mutations into the DNA
9.	Galactosamine ^{[24} ^{7]} (4)	Diffuse type of liver injury simulating viral hepatitis, disrupts the synthesis of essential uridylate nucleotides	Reduces the number of viable hepatocytes, rate of oxygen consumption Tnfα-mediated apoptosis Inflammatory liver injury	Difficulties with reproducibility, refractory anaemia
10.	DEN, NDEA ^[248-249] (4)	Progression of liver fibrosis to HCC	GenerationofROSresultinginoxidativedamagetomacromolecules	Vary with the genetic background, sex, age, other factors
11.	tBHP ^[250] (3)	Oxidative Stress	Inhibition of complex I activity peroxidation of membrane lipids, deplete cellular GSH	
12.	Malathion ^[251] (2)	Cytotoxicity, genotoxicity	Inhibition & accumulation of acetyl cholinesterase activity, leading to the interference with the transmission of nerve impulse	Toxic to other beneficial insect species, highly toxic to aquatic invertebrates
13.	Lead nitrate ^[252] (2)	ProduceOxidativedamagebyenhancingperoxidationofmembranelipids	GSH depletion, elevation in biochemical parameters	Acute intoxication with Pb ²⁺ caused disturbance in the body metabolism
14.	Chronic alcohol ^[253] (1)	Easy to perform Marked elevation of ALT, steatosis	Liver injury, inflammation, fatty liver	Mild steatosis, slight elevation of serum ALT Short-term feeding with no mortality rate, No liver fibrosis

15.	Na ASO ₂ ^[254-255]	Induction of apoptosis,	Activity of 20S	Carcinogenicity
	(1)	inhibition of tumor cell	proteasome, decreased	
		growth	protein expression of	
			PSMB5, SOD1, GPXL	
16.	Cisplatin ^[256]	Solid organ	Accelerate the apoptotic	Yet not clear
	(1)	malignancies,	process, cellular damage	
		Oxidative damage,		
		Increase in the release of		
		pro-inflammatory		
		cytokines		
17.	Bleomycin ^[257] (1	Inhibit fibrosis -	Inflammatory, fibrotic	Molecular alteration
)	interrupting the cell	reactions, induction of pro-	
		cycle	inflammatory cytokines	
18.	Chlorpromazine [[]	Activation of Toll-like	Significant induction of	Interference with
	^{240]} (1)	receptor signalling by	serum TNF) α , prolonged	metabolism
		LPS, LTA	JNK activation	
19.	Cadmium ^[258] (1)	Benign, malignant tumor	Increased activities of	Genetic damage in
		formation	serum hepatic marker	cultured mammalian
			enzymes, level of lipid	cells
			peroxidation indices	
20.	Nimesulide ^[259] (1	Non-steroidal anti-	Preferential inhibitory	Increased risk for
)	inflammatory drug	activity on COX-2 enzyme	hepatotoxicity
21.	Arsenic ^[260]	Apoptosis	Hepatocellular injury,	Carcinogenicity
	(1)		fatty degeneration,	
			progressive fibrosis	
22.	High Fat diet	Induction of liver injury	Higher plasma ALT, AST,	Associated with obesity,
	(HFD) ^[261]		hepatocyte hypertrophy,	diabetes, insulin
	(1)		lipid droplet accumulation,	resistance, steatosis,
			necrosis, inflammatory	steatohepatitis, cirrhosis
			cell infiltration	

DISCUSSION

Liver disease is a worldwide problem. Conventional medicines used in the treatment of liver diseases are sometimes inadequate, may lack efficacy and can develop serious adverse effects. Prevention is a preferred strategy than cure. A series of review papers have been published by *authors* pertaining to liver disorders, therapeutics and medicinal plants used in the treatment of liver diseases with elaborate compilation of classical, proprietary and patented Ayurvedic products ^[2-7]. These reviews also summarised the information about hepatoprotective herbal drugs used in ASU traditional medicine for the treatment of liver diseases.

In this comprehensive review an effort has been made to provide all-inclusive information on liver disorders and potential medicinal plants used for liver diseases in all the three - ASU classical texts, pharmacopoeias and national formularies. Most potential twenty-five medicinal plants are assessed for its phytochemical and pharmacological activities in a comparative and summarised manner for its potentials to develop the safety and effective products ^[262].

Classical or poly herbal formulations

Due to synergism, the best combination of poly-herbal have various types of molecules provide higher activity against a disease to act against a disease complex as in liver dysfunctions by different mechanisms. Several of the large numbers of herbal and herbo-mineral formulations in different dosage forms used for different liver diseases in ASU Systems are studied for their hepatoprotective activities and safety. ASU classical formulations such as *AmalkyadiGhrita*^[263], *Arogyavarghini*^[264], *Emblica officinalis* and Chyavanaprash^[265,266], *AruvadhaChurnam*^[267], *Chara Parpam*^[268], *KadukkaiMaathirai*^[269], *Dawa-Ul-Kurkum*^[270], *Habb-e-Asgand*^[271], and *Sharbat-e- Deenar*^[272]. No adverse effects have been reported with proper administration of designated therapeutic dosages.

Ayurvedic proprietary products like SAL ^[273], Liv 52^[274], Ayush-Liv. 04^[275], *Jigrine^[276]*, *Kabideen Syrup^[277]* and *Patented BV-7310^[278]* have been studied for their hepatoprotective activity against hepatotoxins like carbon tetrachloride, paracetamol, anti-tuberculosis drugs in rat, mice and rabbit models. More than 24 clinical papers and 92 experimental studies on Liv 52, 21 cases of hepatitis and 45 cases of surgery related hepato-biliary disorders treated with an Ayurvedic drug L 2002 (Livotrit) ^[279, 280] and an open study on 10 patients of alcoholic hepatitis with drug Hepafyte ^[281] suggest significant role of poly herbal formulations in liver dysfunctions.

Extracts

Though aqueous, alcoholic, methanol, hydro-alcoholic, petroleum ether and chloroform extracts are most commonly used for testing of 18, 15, 10, 5, 4 and 3 medicinal plants respectively and hydro-methanol, acetone and ether extracts in a plant each. Essentially aqueous extracts of most commonly used *A. paniculata, C. longa, N. sativa* are found to be effective for their different hepatoprotective activities. A traditional recipe of *P. longum* boiled with milk, the ethanol extract of ginger and Cinnamomum bark rich in oil are found to be hepatoprotective. Five different extracts of *A. vera, E. alba, L. echinta* and four different extracts of *G. glabra* have been studied.

Due to high polarity and miscibility with organic solvent water is the most polar solvent used in the extraction of a wide range of polar compounds. It dissolves a wide range of substances. It is cheap, nontoxic, non-flammable, and highly polar. *Alcohol* is also polar in nature, miscible with water, and could extract polar secondary metabolites. *Chloroform* is a nonpolar solvent and is useful in the extraction of compounds such as terpenoids, flavonoids, fats, and oils, while Ether is useful in the extraction of compounds such as alkaloids, terpenoids, coumarins, and fatty acids ^[282-283].

Phytoconstituents

Liver protective plants contain a variety of chemical constituents like phenols, coumarins, lignans, essential oil, monoterpenes, carotinoids, glycosides, flavonoids, organic acids, lipids, alkaloids and xanthenesetc.One or more phytoconstituents of ten plants have been studied in-vivo or in-vitro for hepatoprotective effects. *A. panniculata* tops the list of phytoconstituents with andrographolide (I), andrographiside (II) and neoandrographolide (III), andrographolide and derivatives followed Azadirachtin-A, Nimbolide, Nimbolide and nimbic – 4 from *AI*. Berberine hydrochloride, sanguinarine from *BA* and curcumin from *C.longa* are found to have a variety of hepatoprotective activities both in acute and chronic conditions. Coumestans (wedelolactone and dimethyl wedelolactone extracted from *E. alba* is found to have anti-cytotoxic activity and regenerative activities. Glycyrrhizin and glycyrrhetic acid probably are the most extensively studied phytoconstituents for a variety of and mostly complimentary hepatocytic activities useful in hepatic dysfunction. Picroliv from *kurroa* and phyllanthin from *Phylanthes*species are probably the most known

hepatoprotective plants investigated for viral hepatitis. *Kalunji*, a common dietetic article with medicinal properties has thymoquinone, a studied hepatoprotective. Syringaresinol from *chirayta* a well-known antipyretic plant is also studied for its hepatoprotective activity. Piperine from *P. longum* has added value as a bio-enhancer for its hepatoprotective activity.

Pharmacology models its significance

CCl₄ and APAP also termed paracetamol are the most commonly used experimental models for the evaluation with hepatoprotective and antioxidant activity. Lipid peroxidation is considered a critical factor in the pathogenesis of liver injuries ^[284]. CCL₄ model is widely used because of the inhibition of the radical CCl₃ enervation in the protection against the damage generated, while APAP model is the highly reproducible, dose-dependent hepatotoxicity of APAP and its outstanding translational importance, because acetaminophen overdose is one of the most frequent reasons for ALF in human. Several new animal models are now available to study specific types of liver damage.

Activities

Most of the hepatoprotective plants have antioxidant activity. *Amala* in addition to having NAFLD activity is protective against heavy metal toxicities like one due to Arsenic. The famous immune-modulator *T. cordifolia* is effective against hepatotoxins, particularly due to anti-tuberculosis drugs. *EA* interestingly is found effective in HCV infection and against hepatotoxins (aflatoxins). Curcuma, Cichori, *Kutaki* has distinct activity in NAFLD. Luffa having anti-hyperglycaemic activity could be of interest; strong anti-toxicity activity of Cinnamomum may justify its role as an additive in many formulations.

The activities of the plant drugs could generally be classified as -

- 1. Anti-hepatotoxic- antagonize the effects of any hepato toxins
- 2. Hepatotropic-promote the healing process of the liver
- 3. Hepatoprotective prophylactic-prevent liver affections

It is clearly established that medicinal plants and phytoconstituents can treat chronic liver disease by inhibiting oxidative damage, suppressing fibrogenesis, eliminating virus infection, and preventing or inhibiting tumors' growth. ASU formulations and ingredients not only prevent liver damage but also reverse the damage as evidenced by the scientific studies.

Toxicity

Aloe was surprising reported to be carcinogenic in a single study, use of Berberis, Neem, Cichorium require care with higher doses and long term use the other plants *C.longa, P.kurro, P. zeylanica, T. chebula* have been variably studied for safety. It has been observed that the extractives of several of these plants have been reported to be toxic as in case of *P. zeylanica* root where petroleum ether, acetone and hydro-alcoholic extracts showed Hepatic and renal changes^[285].

CONCLUSIONS

The present review can be concluded as follow-

- 1. Liver diseases in ASU systems have an interesting philosophical background with a long history. The significance of the liver in the context of blood as an important factor for liver dysfunction.
- 2. Natural origin ingredients have been extensively used in ASU traditional medicine for the treatment of liver diseases. Due to synergism, the best combination of poly-herbal provides higher activity by different mechanisms to provide a complete therapy against liver diseases.

- 3. Medicinal plants contain a complex blend of phytochemicals that have the unique ability to address a multiplicity of problems simultaneously. The activities of plants are most likely due to their internal complexity and to the interactions of the different components within the body rather than to one of its specific components.
- 4. Comparatively broad-spectrum benefits and safety as observed in many studies on polyherbal and aqueous extracts as used in traditional medicine do raise questions about use of solvent extracts. Extracts may concentrate one or two isolated constituents which having particular activity but inadvertently overlook some other components that may contribute the same as activity of the whole drug. The issue of standardization could be addressed with advances in analytical methods.
- 5. Need to develop new pre-clinical models to justify integrative approaches to examine specific liver dysfunction is felt to enhance research in liver disorders.
- 6. Ayurveda, Siddha and Unani systems offer highly promising translational opportunities to develop and validate new therapeutics for treatment for liver diseases. The need is to stimulate new research by bridging fundamentals of traditional systems with the advances in biotechnology to explore richness of natural products and ingredients.
- 7. This review will helpto consider new methods for safe and cost-effective formulations not only as hepatoprotective remedies but to better treat serious liver diseases as well.

References

- Mukherjee, P. S., Vishnubhatla, S., Amarapurkar, D. N., Das, K., Sood, A., Chawla, Y. K., Eapen, C. E., Boddu, P., Thomas, V., Varshney, S., Hidangmayum, D. S., Bhaumik, P., Thakur, B., Acharya, S. K., & Chowdhury, A. (2017). Etiology and mode of presentation of chronic liver diseases in India: A multi centric study. *PloS one*, *12*(10), e0187033. https://doi.org/10.1371/journal.pone.0187033
- 2. Bhatt AD, Bhatt NS. Indigenous drugs and liver disease. Indian J Gastroenterol.1996; 15: 63-67
- 3. Valvi AR, Mouriya N, Athawale RB, Bhatt NS. Hepatoprotective Ayurvedic plants a review. J Complement Integr Med. 2016 Sep 1;13(3):207-215. doi: 10.1515/jcim-2015-0110. PMID: 27310002.
- 4. Bhatt N. & Deshpande M. (2018). Int J Ayu Pharm Chem Liver Disorders and Potential Medicinal plants: A Review Study. 2018; 9 (10).10.13140/RG.2.2.15990.45120
- 5. Bhatt N, Deshpande M, Namewar P and Pawar S: A review of classical, proprietary and patented Ayurved products and their ingredients in liver/sleen diseases. Int J Pharm Sci & Res.2018; 910: 4056-70. doi: 10.13040/IJPSR.0975-8232.910.4056-70
- Dr. Narendra Bhatt, Dr. Sudha Revathy, Dr. Manasi Deshpande A Review of Siddha Traditional System Products and Their Ingredients in Liver Diseases, International Journal of AYUSH; 2020: 9 (2);55-74
- Shaikh ZA, Kamyab SS, Deshpande MM, Mulla GJ, Bhatt NS and Moghe AS: A systematic review of Unani formulations for potential in treatment of hepatocellular carcinoma. Int J Pharm Sci & Res 2021; 12(10): 5253-63. doi: 10.13040/IJPSR.0975-8232.12(10).5253-63
- 8. AFI: Ayurvedic Formulary Of India, Vol I-III, 1st Edition Delhi (IN): Government Of India, Ministry Of Health And Family Welfare, Department Of Ayurveda, Yoga & Naturopathy, Unani, Siddha And Homoeopathy.2000, 2003, 2011
- 9. SFI, Siddha Formulary of India, 1st Edition Delhi (IN): Government Of India, Ministry Of Health And Family Welfare, Department of Indian Systems Of Medicine & Homoeopathy. 1992
- 10. UFI: The Unani Pharmacopoeia of India, Part II (Formulations), Vol I-IV, 1st Edition Delhi (IN): Government Of India, Ministry of Health And Family Welfare, Department Of Ayurveda, Yoga & Naturopathy, Unani, Siddha And Homoeopathy.2009,2010,2016,2019

- NFUM: National Formulary of Unani Medicine, Part I-VI, 1st edition Delhi (IN): Government of India, Ministry of Health and Family Welfare, Department of Indian Systems of Medicine & Homoeopathy. 1984,1994, 2001, 2006, 2008, 2011
- API: The Ayurvedic Pharmacopoeia of India, 1st Edition, Part I, Delhi (IN): Government of India, Ministry of Health And Family Welfare, Department of Ayurveda, Yoga & Naturopathy, Unani, Siddha and Homoeopathy.1986, 1999, 2001, 2004; Volume I-VI. doi:10.5455/JREIM.82-142002664006
- 13. API:: The Ayurvedic Pharmacopoeia of India, 1st Edition Part II (Formulations) Delhi (IN): Government of India, Ministry of Health and Family Welfare, Department of Ayurveda, Yoga & Naturopathy, Unani, Siddha and Homoeopathy. 2007,2008,2010,2017; Vol- I-IV
- 14. SPI: Siddha Pharmacopoeia of India, Part I, 1st Edition Delhi (IN): Government of India, Ministry of Health And Family Welfare, Department of Ayurveda, Yoga & Naturopathy, Unani, Siddha and Homoeopathy. 2008,2011; Vol I-II,
- 15. UPI: The Unani Pharmacopoeia of India Part-I. 1st Edition Delhi (IN): Government of India, Ministry of Health And Family Welfare, Department of Ayurveda, Yoga & Naturopathy, Unani, Siddha and Homoeopathy. 2007, 2008, 2009; Vol I-IV
- 16. Essential drug list [EDL], Ayurved, Delhi (IN): Government of India Department of AYUSH, Drug control cell, Ministry of Health and Family Welfare, Department of AYUSH; 2013.
- 17. Essential drug list [EDL], Unani Medicine, Delhi (IN): Government of India Department of AYUSH, Drug control cell, Ministry of Health and Family Welfare, Department of AYUSH; 2013.
- Essential drug list [EDL], Siddha Medicine, Delhi (IN): Government of India Department of AYUSH, Drug control cell, Ministry of Health and Family Welfare, Department of AYUSH; 2013.
- 19. Perspectives of Indian Medicinal Plants in the Management of Liver Disorders, Indian Council of Medical Research; 2008 edition (April 1, 2008)
- 20. Sangeeta Rongson, Niten Barman. A Conceptual Study of Raktavaha Srotas and its Disease. International Journal of Ayurveda and Pharma Research.2017; 5(6):89-95.
- 21. Panda AK, Bhuyan GC, Rao MM. Ayurvedic Intervention for Hepatobiliary Disorders: Current Scenario and Future Prospect. J Tradit Med ClinNatur. 2017; 6:210.
- 22. Pt. Kashinath Pandey, Dr. Gorakhnath Chaturvedi, edited by Pt. Rajeshwardutt Shashtri, Charak Samhita- SavimarshVidyotini Hindivyakhyopeta, Chaukhambha Bharati Academy, Varanasi, Reprint 2008, Vimansthan, chapter 5, verse 7, page 711
- 23. Sushruta, Sushrut Samhita, with commentary of Dalhana, Edited by Vaidya Jadavaji Trikamji, Chaukhambha Orientalia, Varanasi, 8th Edition, 2005, Sutrasthana, Chapter 14, verse 4,page 59
- 24. Lad, Vasant, M.A.Sc., Textbook of Ayurveda Fundamental Principles. 2002; p. 57-58
- 25. Mohamed S. Al-Shinnawy, Ahmed R. Hassan, Dalia A. Ismail and Mohamed A. Shahin The Potential Protective and Therapeutic Effects of Aloe Vera Juice on Malathion Induced Hepatotoxicity in Rabbits The Egyptian Journal of Hospital Medicine. April 2014; Vol. 55, Page 146–158
- 26. H, Tiwari P, Srivastava M, Ghoshal S. Hepatoprotective and Histopathological Activity of Ethanol and Aqueous Extracts of Stem of Aloe Vera Linn (Ghee Gangwar) Against Paracetamol-induced Liver Damage in Rats. International Journal of Pharmacy & Bio-Sciences. 2016 Jan 8
- 27. Chandan BK, Saxena AK, Shukla S, Sharma N, Gupta DK, Suri KA, Suri J, Bhadauria M, Singh B.Hepatoprotective potential of Aloe barbadensis Mill.against carbon tetrachloride induced hepatotoxicity.J Ethnopharmacol. 2007 May; 22; 111(3):560-6. Epub 2007 Jan 14.

- 28. Cui, Yan & Ye, Qing & Wang, Heya & Li, Yingchao & Yao, Weirong & Qian, He. Hepatoprotective Potential of Aloe vera Polysaccharides against Chronic Alcohol-induced Hepatotoxicity in Mice. Journal of the science of food and agriculture. 2014; 94. 10.1002/jsfa.6489
- 29. Guo X, Mei N. Aloe vera: A review of toxicity and adverse clinical effects. *J Environ Sci Health C Environ Carcinog Ecotoxicol Rev.* 2016; 34(2):77–96. doi:10.1080/10590501.2016.1166826
- 30. Patel, Krishna & Gupta, Gajendra & Goyal, Dr. Manoj & Nagori, Badri. Assessment of hepatoprotective effect of Tecomella undulata on paracetamol-induced hepatotoxicity in rats. Revista Brasileira de Farmacognosia. Feb 2011; 21 (1). <u>https://doi.org/10.1590/S0102-695X2011005000020</u>
- 31. D Singh, RS Gupta. Hepatoprotective Activity of Methanol Extract of Tecomella undulata against Alcohol and Paracetamol Induced Hepatotoxicity in Rats. Life Sciences and Medicine Research, Volume 2011: LSMR-26
- 32. M. K. Gole, S. Dasgupta, R. K. Sur & J. Ghosal (1997) *HEPATOPROTECTIVE EFFECT OF AMOORA ROHITUKA*, International Journal of Pharmacognosy, 35:5, 318-322,
- 33. Handa SS, Sharma A., Hepatoprotective activity of andrographolide from Andrographis paniculata against carbontetrachloride. Indian J Med Res 1990, 92, 276-278
- 34. Visen pks, Shukla B, Patnail GK, Dhawan BN, Angrapholide protects rat hepatocytes against paracetamol induced damage. J Ethnopharmacol 1993, 40, 131-136
- 35. A. C. Rana and Y. Avadhoot, Hepatoprotective effects of Andrograhphis paniculata against carbon tetrachloride-induced liver damage," *Archives of Pharmacal Research*, vol. 14, no. 1, pp. 93–95, 1991
- 36. B. R. Choudhury and M. K. Poddar, "Andrographolide and kalmegh (*Andrographis paniculata*) extract: in vivo and in vitro effect on hepatic lipid peroxidation," *Methods and Findings in Experimental and Clinical Pharmacology*, vol. 6, no. 9, pp. 481–485, 1984.
- 37. Choleretic action of andrographolide obtained from *Andrographis paniculata* in rats, Phytotherapy research,1991, <u>Volume5, Issue4</u>, 176-178
- 38. Handa SS, Sharma A., Hepatoprotective activity of andrographolide against galactosamine & Paracetamol intoxication in rats. Indian J Med Res 1990, 92, 284-92
- 39. Dhawan BN Effect of Andraphlide against galactosamine induced hepatotoxic.Fitoterapia 1995,66 415-420 G. S. Tripathi , Y. B. Tripathi
- 40. B. R. Choudhury, S. J. Haque, and M. K. Poddar, In vivo and in vitro effects of kalmegh (Andrographis paniculata) extract and andrographolide on hepatic microsomal drug metabolizing enzymes. Planta Medica. 1987; vol. 53, no. 2, pp. 135–140
- 41. Kapil A, Koul IB, Banerjee SK, Gupta BD. Antihepatotoxic effects of major diterpenoid constituents of Andrographis paniculata, Biochem Pharmacol, 1993, 46 182-185
- 42. Dutta A, Sukul NC.Filaricidal properties of a wild herb, Andrographis paniculata. J.Helminthol 1982, 56 81-84
- 43. Pingale Shirish S, hepatoprotection study of leaves powder of Azadirachta indica a. Juss. International Journal of Pharmaceutical Sciences Review and Research. July – August 2010 ; Volume 3, Issue 2, Article 007
- 44. Kale B. P., Kothekar M. A., Tayade H. P., Jaju J. B., Mateenuddin M. Effect of aqueous extract of Azadirachta indica leaves on hepatotoxicity induced by antitubercular drugs in rats. Indian Journal of Pharmacology. 2003;35:177–180.
- 45. Shivashankaramurthy, K.G. & Kiran, L.J. Evaluation of hepatoprotective activity of aqueous extract of Azadirachta indica (Neem) leaves against paracetamol induced hepatotoxicity in albino rats. Pharmacologyonline. 2011; 2. 1013-1024.

- 46. Bhanwra S. Effect of Azadirachta indica (neem) leaf aqueous extract on paracetamol induced liver damage in rats. Indian Journal of Physiology and Pharmacology. 2000;44(1):64–68.
- 47. Devmurari V. P., Jivani N. P. Hepatoprotective activity of methanolic and aqueous extracts of Azadirchata indica leaves. International Journal of Pharm Tech Research. 2010; 2(2):1037–1040.
- 48. Kalaivani T., Meiganam E., Premkumpatoprotectivear N., Siva R. Studies on hepatoprotective properties of leaf extracts of *Azadirachta indica* A. Juss (Meliaceae) *Ethnobotanical Leaflets*. 2009; 13(2):165–170.
- 49. Mohamed A. Dkhil, Saleh Al-Quraishy, Ahmed M. Aref, Mohamed S. Othman, Kamal M. El-Deib, Ahmed E. Abdel Moneim, The Potential Role of *Azadirachta indica* Treatment on Cisplatin-Induced Hepatotoxicity and Oxidative Stress in Female Rats, 2013; Oxid Med Cell Longev.
- 50. Baligar N. S., Aladakatti R. H., Ahmed M., Hiremath M. B. Evaluation of acute toxicity of neem active constituent, nimbolide and its hepatoprotective activity against acute dose of carbon tetrachloride treated albino rats, International Journal of Pharmaceutical Sciences and Research. 2014;5(8):3455– 3466.
- 51. Baligar N. S., Aladakatti R. H., Ahmed M., Hiremath M. B. Hepatoprotective activity of the neembased constituent azadirachtin-A in carbon tetrachloride intoxicated Wistar rats, Canadian Journal of Physiology and Pharmacology. 2014;92(4):267–277
- 52. Shori Amal Bakr. Evaluation of Acute Toxicity of Water Extract of Azadirachta indica Leaves and Seeds in Rats. Pakistan Journal of Biological Sciences. 2013. 16: 697-700
- 53. Glinsukon, T., R. Somjaree, P. Piyachaturawat and Y. Thebtaranonth. Acute toxicity of nimbolide and nimbic acid in mice, rats and hamsters. Toxicol. Lett., 1986; 30: 159-166
- 54. Wang C, Cao M, Shi DX, Yin ZQ, Jia RY, Wang KY, Geng Y, Wang Y, Yao XP, Yang ZR, Zhao J. A 90-day subchronic toxicity study of neem oil, a Azadirachta indica oil, in mice. Hum Exp Toxicol. 2013 Sep;32(9):904-13. doi: 10.1177/0960327113475677. Epub 2013 Feb 25. PMID: 23444337.
- 55. Gandhi M, Lal R, Sankaranarayanan A, Banerjee CK, Sharma PL, Acute toxicity study of the oil from Azadirachta indica seed (neem oil). J Ethnopharmacol. 1988 May-Jun; 23(1):39-51.
- 56. Ahmad M, Alamgeer, Chaudhary MZ, Nadeem M, Sharif T, Ahmad B. Hepatoprotective effect of Berberis lycium (Royle) in hepatotoxic rabbits. Gomal. Uni. J. Res. 2008; 24:24
- 57. Ahmad M, Alamgeer, Chaudhary MZ, Nadeem M, Sharif T, Ahmad B. Induced Acute Hepatotoxicity in Rats. Asian Journal of Pharmaceutical and Clinical Research.. *Hepatoprotective* effect of Berberis lycium (Royle) in hepatotoxic rabbits. Gomal. Uni. J. Res. 2008; 24:24.
- 58. Gilani AH, Janbaz KH. Preventive and curative effects of Berberis aristata fruit extract on paracetamol and CCl4 induced hepatotoxicity. Phytother Res 1995;9:489–494
- 59. Dehar, N., R. Walia, R. Verma, and P. Pandey. Hepatoprotective Activity of Berberis aristata Root Extract against Chemical 2013; Vol. 6, no. 9, Nov. pp. 53-56,
- 60. Manikyam HK, Ramesh C, Yadav P and Kenttar A: Berberine chloride induced apoptosis in HepG2 cancer cell line *via* PPAR gamma activation and COX2 inhibition by regulating prostaglandin E2 (PGE2) signalling pathway. Int J Pharm Sci & Res 2018; 9(11): 4723-28. doi: 10.13040/IJPSR.0975-8232.9(11).4723-28.
- 61. X. Chang, H. Yan, J. Fei et al., Berberine reduces methylation of the MTTP promoter and alleviates fatty liver induced by a high-fat diet in rats. Journal of Lipid Research, vol. 2010; 51, no. 9, pp. 2504–2515.
- 62. Nitika Singh and Bechan Sharma, Toxicological Effects of Berberine and Sanguinarine. Front Mol Biosci. 2018; 5: 21

- 63. M. T. Olaleye, A. C. Akinmoladun, A. A. Ogunboye, and A. A. Akindahunsi, Antioxidant activity and hepatoprotective property of leaf extracts of *Boerhaavia diffusa* Linn against acetaminophen-induced liver damage in rats, Food and Chemical Toxicology, vol. 48, no. 8-9, pp. 2200–2205, 2010.
- 64. Patel Monali, Verma Ramtej, Hepatoprotective Activity of Boerhavia diffusa Extract, IJPCR, Vol 6, Issue 2, July-September 2014, 233-240
- 65. P. Venkatalakshmi, V. D. Eazhisai, and S. Netaji, Hepatoprotective Activity of Boerhavia diffusa against paracetamol induced toxicity in rats. Journal of Chemical & Pharmaceutical Research. 2011; vol. 3, pp. 229–232
- 66. A. K. S. Rawat, S. Mehrotra, S. C. Tripathi, and U. Shome. Hepatoprotective activity of Boerhaavia diffusa L. roots—a popular Indian ethnomedicine. Journal of Ethnopharmacology. 1997; vol. 56, no. 1, pp. 61–66.
- 67. Beedimani RS, Jeevangi SK. Evaluation of hepatoprotective activity of Boerhaavia diffusa against carbon tetrachloride induced liver toxicity in albino rats. Int J Basic Clin Pharmacol 2015; 4:153-8.
- 68. T. Devaki, K. S. Shivashangari, V. Ravikumar, and P. Govindaraju. Effect of Boerhaavia diffusa on tissue anti-oxidant defense system during ethanol-induced hepatotoxicity in rats. Journal of Natural Remedies. 2005; vol. 5, no. 2, pp. 102–107.
- 69. Nalini G, Ajithadasaruna, Chidambaranathan N and Jegan N. Hepatoprotective activity of hydroalcoholic extract of Boerhaavia diffusa linn against d-galactosamine induced hepatotoxicity in mice. Int J Pharm Sci Res 2018; 9(8): 3367-72. doi: 10.13040/IJPSR.0975-8232. 9(8).3367-72
- 70. Ebere Orish, Orish E Orisakwe, Onyenmechi Johnson, Chudi Emma Dioka, Sub-chronic Toxicity Studies of the Aqueous Extract of Boerhavia diffusa Leaves, 2003, Journal of health science 49(6):444-447
- 71. Singh A, Singh RG, Singh RH, Mishra N, Singh N, An experimental evaluation of possible teratogenic potential in Boerhavia diffusa in albino rats. Planta Med. 1991; 57,315-316
- 72. Gilani AH, Janbaz KH. Evaluation of the liver protective potential of Cichorium intybus seed extract on acetaminophen and CC14-induced damage. Phytomedicine. 1994;1(3):193–197
- 73. Gilani AH, Janbaz KH, Shah BH. Esculetin prevents liver damage induced by paracetamol and CCL4. Pharmacological Research. 1998;37(1):31–35
- 74. Gadgoli C, Mishra SH. Antihepatotoxic activity of Cichorium intybus. Journal of Ethnopharmacology. 1997; 58(2):131–134.
- 75. Souad Elgengaihi, Abdel-Tawab H. Mossa, Amel A. Refaie & Doha Aboubaker, Hepatoprotective Efficacy of Cichorium intybus L. Extract Against Carbon Tetrachloride-induced Liver Damage in Rats. Journal of Dietary Supplements. 2016; Volume 13, Issue 5, Pages 570-584
- 76. Bhaskara Reddy Nallamilli Ch.S.Phani Kumar K.Veer Reddy M.Lakshmi Prasanna V.Maruthi P.Sucharita, Hepatoprotective activity of Cichorium intybus (Linn.) root extract against carbon tetrachloride induced hepatotoxicity in albino Wistar rats, Drug Invention Today. December 2013;Volume 5, Issue 4, Pages 311-314
- 77. Jamshidzadeh, A., Khoshnood, M., Dehghani, Z., Niknahad, H. Hepatoprotective Activity of Cichorium intybus L. Leaves Extract against Carbon Tetrachloride Induced Toxicity. Iranian Journal of Pharmaceutical Research, 2010; Volume 5(Number 1): 41-46
- Zafar R, Mujahid Ali S. Anti-hepatotoxic effects of root and root callus extracts of Cichorium intybus L. Journal of Ethnopharmacology. 1998;63(3):227–231
- 79. Ahmed B, Khan S, Masood MH, Siddique AH. Anti-hepatotoxic activity of cichotyboside, a sesquiterpene glycoside from the seeds of Cichorium intybus. Journal of Asian Natural Products Research. 2008;10 (3-4): 223–231.

- 80. Hepatoprotective effect of Cichorium intybus L, a traditional Uighur medicine, against carbon tetrachloride-induced hepatic fibrosis in rats World J Gastroenterol. 2014; 20(16): 4753–4760.
- Ziamajidi N, Khaghania S, Hassanzadeh G, et al. Amelioration by chicory seed extract of diabetesand oleic acid-induced non-alcoholic fatty liver disease (NAFLD)/non-alcoholic steatohepatitis (NASH) via modulation of PPAR-alpha and SREBP-1. Food and Chemical Toxicology. 2013;58:198– 209
- Physician's desk reference [PDR] for Herbal medicines, Medical Economics company, New Jersy, USA 2000, 181-182
- 83. Eidi A, Mortazavi P, Bazargan M, Zaringhalam J: Hepatoprotective activity of cinnamon ethanolic extract against CCL 4-induced liver injury in rats. EXCLI J. 2012, 11: 495-507.
- 84. El-hadary, Abdalla. Antioxidant and Hepatoprotective effect of Cinnamomum Zeylanicum oil against paracetamol induced hepatotoxicity in male rate. Journal of Agricultural Chemistry and Biotechnology. 2015; 6. 407-418
- 85. Mahyar Dorri, Shirin Hashemitabar & Hossein Hosseinzadeh. Cinnamon (*Cinnamomum zeylanicum*) as an antidote or a protective agent against natural or chemical toxicities: a review, Drug and Chemical Toxicology. 2018; 41:3, 338-351
- 86. Karamalakova, Y. & Nikolova, Galina & Georgiev, Tzvetelin & Gadjeva, Veselina & Tolekova, Anna. Hepatoprotective properties of Curcuma longa L. extract in bleomycin-induced chronic hepatotoxicity. Drug Discoveries & Therapeutics. 2019; 13. 9-16. 10.5582/ddt.2018.01081.
- 87. Singh, I. & Subramaniyan, Vetriselvan & Shankar, J. & Gayathiri, S. & Hemah, C. & Shereenjeet, G. & Yaashini, A. Hepatoprotective activity of aqueous extract of Curcuma longa in ethanol induced hepatotoxicity in albino wistar rats. Int. J. Phytopharmacol. 2012; 3. 226-233
- 88. M.N. Somchit, A. Zuraini, A. Ahmad Bustamam, N. Somchit, M.R. Sulaiman and R. Noratunlina. Protective Activity of Turmeric (Curcuma longa) in Paracetamol-induced Hepatotoxicity in Rats. International Journal of Pharmacology. 2005; 1: 252-256
- 89. Baxla SL, Gora RH, Kerketta P, Kumar N, Roy BK and Patra PH. Hepatoprotective effect of Curcuma longa against lead induced toxicity in Wistar rats. 2013; Vet World 6(9): 664-667, doi: 10.14202/vetworld.2013.664-667
- 90. Hismiogullari, S.E.; Hismiogullari, A.A.; Sunay, F.B.; Paksoy, S.; Can, M.; Aksit, H.; Karaca, O. Yavuz, O. The protective effect of curcumin on carbon tetrachloride induced liver damage. Revue Méd. Vét. 2014, 165, 194–200.
- 91. Eun Jung Kyung et al. Evaluation of Hepatoprotective Effect of Curcumin on Liver Cirrhosis Using a Combination of Biochemical Analysis and Magnetic Resonance-Based Electrical Conductivity Imaging Mediators of Inflammation / 2018, 9 pages. <u>https://doi.org/10.1155/2018/5491797</u>
- 92. Li, B.; Wang, L.; Lu, Q.; Da, W. Liver injury attenuation by curcumin in a rat NASH model: An Nrf2 activation-mediated effect? Ir. J. Med. Sci. 2016, 185, 93–100
- 93. Tokaç, M. Taner, G. Aydın, S. Ozkarde, S. A.B. Dündar, H.Z. Ta, slıpınar, M.Y. Arıkök, A.T. Kılıç, M. Ba, saran, A.A. Basaran, N. Protective effects of curcumin against oxidative stress parameters and DNA damage in the livers and kidneys of rats with biliary obstruction. Food Chem. Toxicol. 2013; 61, 28–35
- 94. Ghoreshi, Z.A. Kabirifar, R. Safari, F. Karimollah, A. Moradi, A. Eskandari-Nasab, E. Hepatoprotective effects of curcumin in rats after bile duct ligation via downregulation of Rac1 and NOX1. Nutrition 2017, 36, 72–78.

- 95. Lin, C.M. Lee, J.F. Chiang, L.L. Chen, C.F. Wang, D. Su, C.L. The protective effect of curcumin on ischemia-reperfusion-induced liver injury in Transplantation Proceedings; Elsevier: Amsterdam, The Netherlands. 2012; Volume 44, pp. 974–977
- 96. Vizzutti, F. Provenzano, A. Galastri, S. Milani, S. Delogu, W. Novo, E.; Caligiuri, A.; Zamara, E. Arena, U.; Laffi, G.; et al. Curcumin limits the fibrogenic evolution of experimental steatohepatitis. Lab. Investig. 2010, 90, 104–115
- 97. Ganiger S, Malleshappa HN, Krishnappa H, Rajashekhar G, Ramakrishna Rao V, Sullivan F.A two generation reproductive toxicity study with curcumin, turmeric yellow, in Wistar rats. Food Chem Toxicol. 2007 Jan;45(1):64-9. E pub 2006 Aug 7.
- 98. Vijayalaxmi, Genetic effects of turmeric and curcumin in mice and rats. Mutat Res. 1980,79,125-132
- 99. Sharma RA, Steward WP, Gescher AJ, Pharmacokinetics and pharmacodynamics of curcumin. Adv Exp Med Biol. 2007; 595:453-70.
- 100. Sreenivasa Rao Damarla, Rajesh Komma, Upendra Bhatnagar, Navin Rajesh, Sadik Mohmad Abdulhamid Mulla. An Evaluation of the Genotoxicity and Subchronic oral Toxicity of Synthetic Curcumin. Journal of Toxicology. Volume 2018, 27 pages
- 101. Mitali Das, Alaka Das, Evaluation of Hepatoprotective Activity of Cuscuta Reflexa Roxb.Stem On Experimental Animals. Indian Journal of Applied Research. 2017; Volume, Issue.
- 102. Amaresh, Panda & Seemanchala, Rath & Debashis, Pradhan & Mahanty, Arpan & Kumar, Gupta & Bala, Nripendra. Hepatoprotective Activity of Whole Part of the Plant Cuscuta reflexa Roxb. (Convolvulaceae) in Chloroform, Ethanol and Paracetamol Induced Hepatotoxic Rat Models. International Journal of Pharmaceutical and Clinical Research. 2014;6. 127-132
- 103. Shweta Mishra, Niraj Dixit, Investigations on hepatoprotective effect of extract in Paracetamol Cuscuta reflexa induced toxicity Asian Journal of Pharmacy and Pharmacology 2019; 5(4):750-754
- 104. Nishant Singh Katiyar, Amrit Pal Singh , Saravanan K , Anil Kumar Gangwar , N. Venkat Rao, Evaluation of hepatoprotective activity of stem extracts of Cuscuta reflexa (roxb) on thioacetamide induced liver damage in rats, World J Pharm Sci 2015; 3(5): 872-877
- 105. Arun K, Balasubramanian U., Comapartive Study on Hepatoprotective activity of Phyllanthus amarus and Eclipta prostrata against alcohol induced in albino rats. International journal of environmental sciences 2011, 2(1), 361-379.
- 106. A. K. Saxena, B. Singh, K. K. Anand. Hepatoprotective effects of Eclipta alba on subcellular levels in rats. Journal of Ethnopharmacology. 1993. vol. 40, no. 3, pp. 155–161.
- 107. B. Singh, A. K. Saxena, B. K. Chandan, S. G. Agarwal, K. K. Anand, In vivo hepatoprotective activity of active fraction from ethanolic extract of Eclipta alba leaves. Indian Journal of Physiology and Pharmacology. 2001; vol. 45, no. 4, pp. 435–441.
- 108. B. Singh, A. K. Saxena, B. K. Chandan, S. G. Agarwal, M. S. Bhatia, and K. K. Anand, "Hepatoprotective effect of ethanolic extract of Eclipta alba on experimental liver damage in rats and mice," Phytotherapy Research, vol. 7, no. 2, pp. 154–158, 1993.
- R. Zafar and B. P. S. Sagar. Hepatoprotective and cardiac inhibitory activities of ethanolic extracts from plant leaves and leaf callus of Eclipta alba. Pharmaceutical Biology. 2000. vol. 38, no. 5, pp. 357–361.
- T. Thirumalai, E. David, V. Therasa, E. K. Elumalai. Restorative effect of Eclipta alba in CCl4 induced hepatotoxicity in male albino rats. Asian Pacific Journal of Tropical Disease. 2011; vol. 1, no. 4, pp. 304–307.

- Prabu K, Kanchana N, Sadiq AM., Hepatoprotective effect of Eclipta alba on paracetamol induced liver toxicity in rats. Journal of Microbiology and Biotechnology Research. 2011, 1 (3), 75-79.
- 112. S. R. Parmar, P. H. Vashrambhai, and K. Kalia. Hepatoprotective activity of some plants extract against paracetamol induced hepatotoxicity in rats. Journal of Herbal Medicine and Toxicology.2010; vol. 4, pp. 101–106.
- 113. K. Kumar, A. K. Katiyar, M. Swamy, Y. P. Sahni, and S. Kumar. Hepatoprotective effect of Eclipta alba on experimentally induced liver damage in rats. Indian Journal of Veterinary Pathology. 2013; vol. 37, pp. 159–163.
- 114. Mogre K, Vora KK, Sheth UK. Effect of Picrorhiza kurro and Eclipta alba on Na+/K+ATPase in hepatic injury by hepatotoxic agenta. Indian JPharmacol. 1981, 13 253-259
- 115. V. N. Murthy, B. P. Reddy, V. Venkateshwarlu, and C. K. Kokate, Antihepatotoxic activity of *Eclipta alba, Tephrosia purpurea* and *Boerhaavia diffusa, Ancient Science of Life*. 1992; vol. 11, pp. 182–186,
- 116. Lin SC, Yao CJ, Lin CC, Lin YH. Hepatoprotective Activity of Taiwan Folk Medicine: Eclipta prostrata Linn against Various Hepatotoxins Induced Acute Hepatotoxicity. Phytotherapy Research 1998, 10(6), 483 490.
- 117. V. K. Lal, A. Kumar, P. Kumar, and K. S. Yadav, Screening of leaves and roots of Eclipta alba for hepatoprotective activity. Archives of Applied Science Research. 2010; vol. 2, no. 1, pp. 86-94
- 118. H. Wagner, B. Geyer, Y. Kiso, H. Hikino, and G. S. Rao. Coumestans as the main active principles of the liver drugs Eclipta alba and Wedelia calendulacea. Planta Medica. 1986; vol. 5, pp. 370–374.
- 119. D. Manvar, M. Mishra, S. Kumar, and V. N. Pandey. Identification and evaluation of anti hepatitis C virus phytochemicals from *Eclipta alba*. *Journal of Ethnopharmacology*. 2012; vol. 144, no. 3, pp. 545–554
- 120. Thilakchand KR, Mathai RT, Simon P, Ravi RT, Baliga-Rao MP, Baliga MS. Hepatoprotective properties of the Indian gooseberry (Emblica officinalis Gaertn): a review. Food Funct. 2013 Oct;4(10):1431-41. doi: 10.1039/c3fo60237k. PMID: 23978895
- 121. Damodara Reddy V, Padmavathi P, Gopi S, Paramahamsa M, Varadacharyulu NCh. Protective effect of Emblica officinalis against alcohol-induced hepatic injury by ameliorating oxidative stress in rats. Indian J Clin Biochem 2010; 25:419-24.
- 122. Reddy VD, Padmavathi P, Varadacharyulu NCh. Emblica officinalis protects against alcohol-induced liver mitochondrial dysfunction in rats. J Med Food 2009; 12:327-33.
- 123. Sharma A, Sharma MK, Kumar M. Modulatory role of Emblica officinalis fruit extract against arsenic induced oxidative stress in Swiss albino mice. Chem Biol Interact 2009;180:20-30.
- 124. Mir AI, Kumar B, Tasduq SA, Gupta DK, Bhardwaj S, Johri RK. Reversal of hepatotoxin-induced pre-fibrogenic events by Emblica officinalis A histological study. Indian J Exp Biol 2007; 45:626-9.
- 125. Sultana S, Ahmed S, Sharma S, Jahangir T. Emblica officinalis reverses thioacetamide-induced oxidative stress and early promotional events of primary hepatocarcinogenesis. J Pharm Pharmacol 2004; 56:1573-9.
- 126. Tasduq SA, Mondhe DM, Gupta DK, Baleshwar M, Johri RK. Reversal of fibrogenic events in liver by Emblica officinalis (fruit), an Indian natural drug. Biol Pharm Bull. 2005; 28:1304-6.
- 127. Sultana S, Ahmad S, Khan N, Jahangir T. Effect of Emblica officinalis (Gaertn) on CCl4 induced hepatic toxicity and DNA synthesis in Wistar rats. Indian J Exp Biol 2005;43:430-6

- 128. Tasduq SA, Kaisar P, Gupta DK, Kapahi BK, Maheshwari HS, Jyotsna S, et al. Protective effect of a 50% hydroalcoholic fruit extract of Emblica officinalis against anti-tuberculosis drugs induced liver toxicity. Phytother Res 2005; 19:193-7.
- 129. Malar H. L. Bai SMM, Hepatoprotective activity of Phyllanthus emblica against paracetamol induced hepatic damage in wistar albino rats. African Journal of Basic and Applied science. 2009;[1-2], 21-25
- Gulati RK, Agarwal S, Hepato protective studies on Phtllanthus emblica Linn and quercetin, Indian Journal of Experimental Biology. 1995. 33 [4] ,261-8
- 131. Chen KH, Lin BR, Chien CT, Ho CH. Emblica officinalis Gaertn attentuates N-nitrosodiethylamine-induced apoptosis, autophagy, and inflammation in rat livers. J Med Food 2011;14:746-55
- Jeena KJ, Joy KL, Kuttan R. Effect of Emblica officinalis, Phyllanthus amarus and Picrorrhiza kurroa on N-nitrosodiethylamine induced hepatocarcinogenesis. Cancer Lett. 1999 Feb 8;136(1):11-6. doi: 10.1016/s0304-3835(98)00294-8. PMID: 10211933.
- 133. Hai Zhong Huo, Bing Wang, Yong Kang Liang, Yong Yang Bao, Yan Gu. Hepatoprotective and Antioxidant Effects of Licorice Extract against CCl₄-Induced Oxidative Damage in RatsInt J Mol Sci. 2011; 12(10): 6529–6543.
- 134. Yin, Guojun & Cao, Liping & Xu, Pao & Jeney, Galina & Nakao, Miki & Lu, Chengping. Hepatoprotective and antioxidant effects of Glycyrrhiza glabra extract against carbon tetrachloride (CCl₄)-induced hepatocyte damage in common carp (Cyprinus carpio). Fish physiology and biochemistry. 2011; 37. 209-16.
- 135. Sharma V, Agrawal RC (2014) In vivo antioxidant and hepatoprotective potential of Glycyrrhiza glabra extract on carbon tetra chloride (CCl4) induced oxidative stress mediated hepatotoxicity. Int J Res Med Sci 2:314–320
- 136. Abd-Al-Sattar L and Laylani S. Hepatoprotective effect of Glycyrrhiza glabra L. extracts against carbon tetrachloride-induced acute liver damage in rats. International Journal of Veterinary Science, Medicine & Research 2016; 1(1): 1-8.
- 137. Al-Razzuqi RAM, Al-Jawad FH, Al-Hussaini JA, Al-Jeboori AA. Hepato-protective effect of Glycyrrhiza glabra in carbon tetrachloride-induced model of acute liver injury. J Phys Pharm Adv 2012; 2(7): 259-263
- 138. Moon A and Kim SH. Effect of Glycyrrhiza glabra roots and glycyrrhizin on the glucuronidation in rats. Planta Med 1997; 63(2):115-119
- 139. Sato H, Goto W, Yamamura J, Kurokawa M, Kageyama S, Takahara T, Watanabe A, Shiraki K. Therapeutic basis of glycyrrhizin on chronic hepatitis B. Antivir Res. 1996; 30:171–177
- 140. Van Rossum TG, Vulto AG, De Man RA, Brouwer JT, Schalm SW. Glycyrrhizin as a potential treatment of chronic hepatitis C. Aliment Pharmacol Ther. 1998; 12:199–205
- 141. Jeong HG, You HJ, Park SJ, Moon AR, Chung YC, Kang SK, Chun HK. Hepatoprotective effects of 18β-glycyrrhetinic acids on carbon tetrachloride-induced liver injury, inhibition of cytochrome P450 2E1 expression. 2002; Pharmacol Res 46(3):221–227
- 142. Tu CT, Li J, Wang FP, Li L, Wang JY, Jiang W, Glycyrrhizin regulates CD4+T cell response during liver fibrogenesis via JNK, ERK and PI3K/AKT pathway. Int Immunopharmacol. 2012 Dec; 14(4):410-21
- 143. Sato H, Kageyama S, Yamamoto H, et al. Glycyrrhizin renders cells resistant to apoptosis induced by human and feline immunodeficiency virus. Journal of Traditional Medicines. 2011; 28(3):139–148.

- 144. Abe N, Ebina T, Ishida N. Interferon induction by glycyrrhizin and glycyrrhetinic acid in mice. Microbiology and Immunology. 1982; 26(6):535–539.
- 145. Epstein MT.Espiner EA,Donald RA, Hughes H. Effect of eating liquorice on the reninangiotensin alsosterone axis in normal subjects. Br Med J. 1977;1,488-490
- 146. Bahar Ahmed, Tanveer Alam, Shah A Khan, Hepatoprotective Activity Of Luffa EchinataFruits,Journal Of Ethnopharmacology, Volume 76, Issue 2, 2001; Pages 187-189
- P Lauria, VN Sharma, S Vanjani, BC Sangal. The Effect of Luffa Echinata in Liver Injury. Fnd. J. Pharmac. 1976; 8 (2) 129-L 33.
- 148. Jakhmola V, Pawar VK, Lal VK. Hepatoprotective Effect of Acetone Extract Of Luffa Echinata Root Against Carbon Tetrachloride Induced Liver Injury In Rats. Pharmacologyonline 2010; 1: 849-855
- 149. Bapat SK, Chandra V. The effect of Luffa echinata (Roxb) on experimental jaundice in rats. Indian J Physiol Pharmacol 1968; 12(3): 119-120.
- Richa Dogar, Dr. S. K. Yadav, Dr. B.P. Nagori, Dr. K. Mathur, Dr. M. Goyal. Hypoglycaemic Effect of Luffa Echinata In Alloxan Induced Diabetic Rats. Indian Journal of Drugs. 2018; 6(4), 201-217
- 151. Toppo R, Roy BK, Gora RH, Baxla SL, Kumar P. Hepatoprotective activity of Moringa oleifera against cadmium toxicity in rats. Vet World. 2015;8(4):537-540. doi:10.14202/vetworld.2015.537-540
- 152. Abdel Fattah ME, Sobhy HM, Reda A, Abdelrazek HMA. Hepatoprotective effect of Moringa oleifera leaves aquatic extract against lead acetate-induced liver injury in male Wistar rats. Environ Sci Pollut Res Int. 2020 Dec;27(34):43028-43043. doi: 10.1007/s11356-020-10161-z. Epub 2020 Jul 28. PMID: 32725563.
- 153. Pari L, Kumar NA. Hepatoprotective activity of Moringa oleifera on antitubercular druginduced liver damage in rats. J Med Food. 2002 Fall;5(3):171-7
- 154. Aly, O., Abouelfadl, D.M., Shaker, O.G. et al. Hepatoprotective effect of Moringa oleifera extract on TNF-α and TGF-β expression in acetaminophen-induced liver fibrosis in rats. 2020; Egypt J Med Hum Genet 21, 69.
- 155. Mbagwu, Herbert, EtokNsikan, &EkpoEmem. Hepatoprotective activity of the methanolic leaf extract of Moringa oleifera (Lam) against chemically-induced Liver Toxicity. *International Journal of Pharmacology and Toxicology [Online]*, 4.1 (2016): 40-46
- 156. Farrag AR, Mahdy KA, Abdel Rahman GH, Osfor MM. Protective effect of Nigella sativa seeds against lead-induced hepatorenal damage in male rats. Pak J Biol Sci. 2007 Sep 1;10(17):2809-16. doi: 10.3923/pjbs.2007.2809.2816. PMID: 19090181.
- 157. Chatterjee M, Sil PC. Hepatoprotective effect of aqueous extract of Phyllanthus niruri on nimesulide-induced oxidative stress in vivo. Indian J Biochem Biophys. 2006 Oct;43(5):299-305. PMID: 17133737.
- 158. Renuka MR, Rahim S. Hepatoprotective Effect of Phyllanthus Niruri Aqueous Extract on Paracetamol Induced Stress in the Fish Anabas Testudineu , International Journal of Information Research and Review. November, 2017; Vol. 04, Issue, 11, pp.4705-4707
- 159. Kanter M, Coskun O, Budancamanak M, Hepatoprotective effects of Nigella sativa L and Urtica dioica L on lipid peroxidation, antioxidant enzyme systems and liver enzymes in carbon tetrachloride-treated rats.World J Gastroenterol. 2005 Nov 14; 11(42):6684-8.

- 160. Yildiz F, Coban S, Terzi A, Ates M, Aksoy N, Cakir H, Ocak AR, Bitiren M. Nigella sativa relieves the deleterious effects of ischemia reperfusion injury on liver.World J Gastroenterol. 2008 Sep 7; 14(33):5204-9.
- 161. El-Sayed WM. Upregulation of chemoprotective enzymes and glutathione by *Nigella sativa* (black seed) and thymoquinone in CCl4-intoxicated rats. Int J Toxicol. 2011; 30:707–714.
- 162. Nagi MN, Almakki HA, Sayed-Ahmed MM, Al-Bekairi AM. Thymoquinone supplementation reverses acetaminophen-induced oxidative stress, nitric oxide production and energy decline in mice liver. Food Chem Toxicol. 2010;48:2361–2365
- 163. Talib WH, Abukhader MM. Combinatorial effects of thymoquinone on the anti-cancer activity and hepatotoxicity of the prodrug CB 1954. Sci Pharm. 2013; 81:519–530.
- 164. Aycan IO, Tufek A, Tokgoz O, Evliyaoglu O, Firat U, Kavak GO, et al. Thymoquinone treatment against acetaminophen-induced hepatotoxicity in rats. Int J Surg. 2014; 12:213–218.
- 165. Sayed-Ahmed, Mohamed M et al. Thymoquinone attenuates diethylnitrosamine induction of hepatic carcinogenesis through antioxidant signaling. Oxidative medicine and cellular longevity vol. 3, 4 (2010): 254-61. doi:10.4161/oxim.3.4.12714
- 166. Al-Ali A, Alkhawajah AA, Randhawa MA, Shaikh NA, Oral and intraperitoneal LD50 of thymoquinone, an active principle of Nigella sativa, in mice and rats.J Ayub Med Coll Abbottabad. 2008 Apr-Jun; 20(2):25-7.
- 167. Khader M, Bresgen N, Eckl PM, In vitro toxicological properties of thymoquinone., Food Chem Toxicol. 2009 Jan; 47(1):129-33
- 168. Zaoui A, Cherrah Y, Mahassini N, Alaoui K, Amarouch H, Hassar M, Acute and chronic toxicity of Nigella sativa fixed oil.Phytomedicine. 2002 Jan; 9(1):69-74
- 169. Wongnawa, M., Thaina, P., Bumrungwong, N., Nitiruangjarat, A., Muso, A., Prasartthong, V. 2005. Congress on Medicinal and Aromatic Plants. Vol. 6: Traditional Medicine and Nutraceuticals. ISHS Acta Horticulturae 680. <u>10.17660/ActaHortic.2005.680.30</u>
- 170. Surya Narayanan, B., Latha, P., Rukkumani, R. Protective effects of Phyllanthus amarus on fibrotic markers during alcohol and polyunsaturated fatty acid-induced toxicity. Toxicology Mechanisms and Methods 2011; 21, 48–52
- 171. Syed Asad B, Iqbal MM, Kiranmai M, Ibrahim M. Hepatoprotective Activity of Phyllanthus Amarus Seeds Extracts in CCl4 Treated Rats: In vitro & In Vivo. Global Journal of Medical Research.2012; 12(6):39-49.
- 172. Visweswaram, D., Rao, P.R., Satyanarayana, S., 1994. A noninvasive method for screening hepatoprotective drugs against carbon tetrachloride-induced hepatotoxicity. Indian Journal of Pharmacology 26, 301–303.
- 173. Krithika, A., Verma, R.R.J., 2009b. Mitigation of carbon tetrachloride induced damage by Phyllanthus amarus in liver of mice. Acta Poloniae Pharmaceutica Drug Research 66, 439–445.
- 174. Krithika, A.R., Verma, R.J., 2009a. Ameliorative potential of Phyllanthus amarus against carbon tetrachloride induced hepatotoxicity. Acta Poloniae Pharmaceutica – Drug Research 66, 579– 583
- 175. Naaz, F., Javed, S., Abdin, M.Z., 2007. Hepatoprotective effect of ethanolic extract of Phyllanthus amarus Schum.et.Thonn.on afltoxin B1-induced liver damage in mice. Journal of Ethnopharmacology 113, 503–509
- 176. Zahra A. Amin, Mehmet Bilgen, Mohammed A. Alshawsh, Hapipah M. Ali, A. Hamid A. Hadi, and Mahmood A. AbdullaProtective Role of *Phyllanthus niruri* Extract against Thioacetamide-

Induced Liver Cirrhosis in Rat Model, Evidence-Based Complementary and Alternative Medicine. 2012

- Faremi, T.Y, Suru, S.M., Fafunso, M.A., Obioha, U.E. Hepatoprotective potentials of Phyllanthus amarus against ethanol induced oxidative stress in rats. Food and Chemical Toxicology. 2008; 46, 2658–2664
- 178. Joshi, C.S., Priya, E.S. Glucuronidase inhibitory effect of phenolic constituents from Phyllanthus amarus. Pharmaceutical Biology. 2007; 45, 363–365.
- 179. Ezzat MI, Okba MM, Ahmed SH, El-Banna HA, Prince A, Mohamed SO, et al. In-depth hepatoprotective mechanistic study of *Phyllanthus niruri*: In vitro and in vivo studies and its chemical characterization. PLoS ONE.2020; 15(1)
- 180. Bhattacharjee R, Sil PC. Protein isolate from the herb, Phyllanthus niruri, protects liver from acetaminophen induced toxicity. Biomedical Research. 2006; 17(1):75-79.
- 181. Sabir SM, Rocha JBT. Water extractable phytochemical from Phyllanthus niruri exhibit distinct in vitro antioxidant and in vivo hepatoprotective activity against paracetamol induced liver damage in mice. Food chemistry, 2008; 111:845-851
- 182. Pramyothin P, Ngamtin C, Poungshompoo S, Chaichantipyuth C. Hepatoprotective activity of Phyllanthus Amarus Schum. Thonn extract in ethanol treated rats: in vitro and in vivo studies. Journal of Ethnopharmacology 2007; 114(2):169-173
- 183. Praveen Kumar and Shukla, S.K. Hepatoprotective Efficacy of Picrorhiza kurroa in experimentally induced Hepatotoxicity in Cockerels. Int.J.Curr.Microbiol.App.Sci. 2017; 6(4): 2614-2622
- 184. Sinha S, Bhat J, Joshi M, Sinkar V, Ghaskadbi S. Hepatoprotective activity of Picrorhiza kurroa Royle Ex. Benth extract against alcohol cytotoxicity in mouse liver slice culture. Int J Green Pharm 2011;5:244-53
- 185. Sapna N. Shetty, Sushma Mengi, Rama Vaidya, and Ashok D. B. VaidyaA study of standardized extracts of *Picrorhiza kurroa* Royle ex Benth in experimental nonalcoholic fatty liver disease, J Ayurveda Integr Med. 2010 Jul-Sep; 1(3): 203–210.
- 186. Rastogi R, Saksena S, Garg NK, Kapoor NK, Agarwal DP, Dhawan BN. Picroliv protects against alcohol-induced chronic hepatotoxicity in rats. Planta Med 1996;62:283-5.14
- 187. Visen PK, Shukla B, Patnaik JK. Hepatoprotective activity of Picroliv. The active principle of Picrorhiza kurroa on rat hepatocytes against paracetamol toxicity. Drug Dev Res 1991;22:209-10
- 188. Shukla B, Visen PK, Patnaik GK, Dhawan BN. Choleretic effect of picroliv, the hepatoprotective principle of Picrorhiza kurroa. Planta Med 1991; 57:29-33.15.
- 189. Dwivedi Y. Rastogi R. Mehtorta R Garg NK, Dhawan BN, Picoliv protects against aflatoxin B1 acute hepatotoxicity in rats. Pharmacol Res 1993,27 189-199
- 190. Ravi Rastogi, Seema Saksena, N. K. Garg, B. N. Dhawan Effect of picroliv on antioxidantsystem in liver of rats, after partial hepatectomy. Phytotherapy research. August 1995;Volume 9, Issue5, "Pages 364-367
- 191. Praveen C. Verma, Vaishali Basu, Vijayta Gupta, Gauri Saxena, Laiq Ur Rahman. Pharmacology and Chemistry of a Potent Hepatoprotective Compound Picroliv Isolated from the Roots and Rhizomes of Picrorhiza kurroa Royle ex Benth. (Kutki), Current Pharmaceutical Biotechnology. 2009;Volume 10 (6).
- 192. Singh M, Tiwari V, Jain A, Ghoshal *S*. Protective activity of picroliv on hepatic amoebiasis associated with carbon tetrachloride toxicity. Indian J Med Res. 2005 May; 121(5):676-82.

- 193. Chander R, Dwivedi Y, Rastogi R, Sharma SK, Garg NK, Kapoor NK, Dhawan BN, Evaluation of hepatoprotective activity of picroliv (from Picrorhiza kurroa) in Mastomys natalensis infected with Plasmodium berghei, Indian J Med Res. 1990 Feb;92:34-7.
- 194. Chander R, Kapoor NK, Dhawan BN. Picroliv affects gamma-glutamyl cycle in liver and brain of Mastomys natalensis infected with Plasmodium berghei. Indian J Exp Biol. 1994 May; 32(5):324-7.
- 195. Chander R, Kapoor NK, Dhawan BN, Effect of picroliv on glutathione metabolism in liver and brain of Mastomys natalensis infected with Plasmodium berghei. Indian J Exp Biol. 1992 Aug; 30(8):711-4.
- 196. Anasari RA. et al. Hepatoprotective activity of Kutkin- the iridoid glycoside mixture of Picrorhiza kurrooa. Indian J Med Res. 1988;87,401-404
- 197. Patel, Jagruti & SHAH, Urvi. (2009). Hepatoprotective activity of Piper longum traditional milk extract on carbon tetrachloride induced liver toxicity in Wistar rats. Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas. 8.
- 198. Kesarwani K, Gupta R, Mukerjee A. Bioavailability enhancers of herbal origin: an overview. Asian Pac J Trop Biomed. 2013; 3(4):253–266.
- 199. O Abdel Salam, S Nofal, S El-Shenawy, N Shaffie. Effect of Piperine on Liver Damage and Bone Changes Caused By Bile Duct Ligation In Rats. The Internet Journal of Pharmacology.2007 Volume 5 Number 2.
- 200. Mananvalan G and Singh J. Chemical and some pharmacological studies on leaves of P.longum Linn., Indian J. Pharm.Sci 1979; 41:190
- 201. Pawinee PiyachaturawatThirayudh GlinsukonChaivat Toskulkao, Acute and subacute toxicity of piperine in mice, rats and hamsters, Toxicology Letters. 1983;Volume 16, Issues 34, Pages 351-359
- 202. Tayubi IA, Desai A, Madar IH, Al Sadh H. Hepatoprotective activity of plumbago zeylanica linn. Against carbon tetrachloride induced heaptotoxicity in rats. Int J Sci Innovs, 2018; 5 (2): 094-098.
- 203. Kumar R, Kumar S, Patra A, Jayalakshmi S (2009). Hepatoprotective activity of aerial parts of Plumbago zeylanica linn against carbon tetrachloride-induced hepatotoxicity in rats. Int. J. Pharma. Pharm. Sci. 2009; 1: 171-175
- 204. N. Kanchana And A. Mohamed Sadiq, Hepatoprotective Effect Of Plumbago Zeylanica On Paracetamol Induced Liver Toxicity In Rats, International Journal of Pharmacy and Pharmaceutical Sciences. 2011; Vol 3, Issue 1.
- 205. Kumar Patil, Roy S.Kholkute SD, Hegde HV, Nair V. Comparative toxicity profiles of Pulmbago zeylanica L. root pterolleum ether, acetone and hydroalcoholic extracts in Wistar rats. AYU 2015:36:329-34
- 206. Nagalekshmi R., Menon A., Chandrasekharan D. K., Nair C. K. K. Hepatoprotective activity of Andrographis paniculata and Swertia chirayita. Food Chem. Toxicol. 2011; 49:3367–3373.
- 207. Karan M, Vasisht K, Handa S.S., Antihepatotoxic activity of Swertia chirata on paracetamol and galactosamine induced hepatotoxicity in rats. Phytotherapy Research. 1999; 13(2), 95-01
- 208. Chakravaty AK, Mukhopadhyay S. Moitra SK, Das B Syringaresinol, a hepatoprotective agent and other constituents fom Swertia chirayita. Indian J. Chem. 1994; 33B 405-408
- 209. Mukherjee S, Sur A, Maiti B.R., Hepatoprotective effect of Swertia chirata on rat. Indian J Exp Biol., 1997, 35(4), 384-8.

- Gunjegaonkar SM, Kshirsagar SS, Bayas JP. Hepatoprotective and Antioxidant potential of Tephrosia Purprea in Paracetamol Induced Hepatotoxicity. Int J of Pharmc Res [Internet]. 2016 Mar. 30; 6(3):104-8.
- 211. Shah et al Evaluatioof Hepatoprotective Activity of Ethyl Acetate Fractio of Tephrosia Purpurea. Pharmacologyonline. 2011; 3: 188-194
- 212. Ravuri Halley Gora, Sushma Lalita Baxla, Priscilla Kerketta, Subhasree Patnaik, Birendra Kumar Roy, Hepatoprotective activity of *Tephrosia purpurea* against arsenic induced toxicity in rats, Indian J Pharmacol. 2014 Mar-Apr; 46(2): 197–200. doi: 10.4103/0253-7613.129317
- 213. Khatri A, Garg A, Agrawal SS Evaluation of hepatoprotective activity of aerial parts of Tephrosia purpurea L. and stem bark of Tecomella undulata, Journal of Ethnopharmacology, 18 Nov 2008, 122(1):1-5
- 214. Tasduq, S.A., K. Singh, N.K. Satti, D.K. Gupta, K.A. Suri and R.K. Johri. *Terminalia chebula* (fruit) prevents liver toxicity caused by sub-chronic administration of rifampicin, isoniazid and pyrazinamide in combination. Hum. Exp. Toxicol. 2006; 25: 111
- 215. Srisesharam Srigopalram, Indira A Jayraaj, Effect of Terminalia Chebula Retz on Den Induced Hepatocellular Carcinogenesis In Experimental Rats, International Journal of Pharmacy and Pharmaceutical Sciences. 2012. 4(2).
- 216. Min-Kyung Choi, Hyeong-Geug Kim Jong-Min Han, Jin-Seok Lee, Jong Suk Lee, Sun Ho Chung, and Chang-Gue Son, Hepatoprotective Effect of *Terminalia chebula* against *t*-BHP-Induced Acute Liver Injury in C57/BL6 Mice. Evidence-Based Complementary and Alternative Medicine. 2015
- 217. Tania Yeasmin, Qazi Shamima Akhter, Syeda Tasfia Siddika, Fayeza Karim. Effect of Terminalia Chebula (Haritaki) on Serum Aspartate Aminotransferase, Alanine Aminotransferase in Paracetemol induced liver damage in Wister Albino Rats, J Bangladesh Soc Physiol. 2015, June; 10(1): 1-5
- 218. V, B., and L. T. Hepatoprotective Activity Of Ethanolic Extract Of Terminalia Chebula Fruit Against Ethanol Induced Hepatotoxicity In Rats". Asian Journal of Pharmaceutical and Clinical Research, Vol. 10, no. 11, Nov. 2017, pp. 55-58,
- 219. Lee H.-S., Jung S.-H., Yun B.-S., Lee K.-W.Isolation of chebulic acid from *Terminalia chebula* Retz.and its antioxidant effect in isolated rat hepatocytes. Archives of Toxicology. 2007;81(3):211–218
- 220. Hussein R. H., Khalifa F. K. The protective role of ellagitannins flavonoids pretreatment against *N*-nitrosodiethylamine induced-hepatocellular carcinoma. *Saudi Journal of Biological Sciences*. 2014; 21(6):589–596.
- 221. Rathore, H. A Study on the Cytological Effects of Myrobalan (Fruit of Terminalia chebula) in Allium Tests. *Ethnobotanical Leaflets*, 2006, 9.
- 222. Arora, E. Brits, S. Kaur, K. Kaur, R.S. Sohi, S. Kumar, L. Verschaeve L. Evaluation of genotoxicity of medicinal plant extracts by the Comet and VITOTOX tests. J. Environ. Pathol.Toxicol.Oncol. 2005; 24(3): 193-200.
- 223. Stance MH, A Nagy A., Tosa M., Vlad L. Hepatoprotective effect of orally administered melatonin & Tinospora cordifolia in experimental jaundice. Chirurgia (bucur) 2011; 106(2), 205-10
- 224. PEER F, Sharma PC. Tinospora Cordifolia in CCl₄ induced Hepatopathy in Goats. Indian Journal Vet. Med 1989; 9 154-156

- 225. Sharma B, Dabur R. Protective Effects of Tinospora cordifolia on Hepatic and Gastrointestinal Toxicity Induced by Chronic and Moderate Alcoholism. Alcohol Alcohol. 2016 Jan; 51(1):1-10. doi: 10.1093/alcalc/agv130. Epub 2015 Nov 19. PMID: 26589585.
- 226. Nagarkatti DS1, Rege NN, Desai NK, Dahanukar SA. Modulation of Kupffer cell activity by Tinospora cordifolia in liver damage. J Postgrad Med. 1994; Apr-Jun;40(2):65-7.
- 227. B. T. Kavitha, S. D. Shruthi, S. Padmalatha Rai, and Y. L. RamachandraPhytochemical analysis and hepatoprotective properties of *Tinospora cordifolia* against carbon tetrachloride-induced hepatic damage in rats, J Basic Clin Pharm. June -August 2011; 2(3): 139–142.
- 228. Kumar, V. & Modi, P.K. & Saxena, K.K.Exploration of hepatoprotective activity of aqueous extract of Tinospora cordifolia - An experimental study. Asian Journal of Pharmaceutical and Clinical Research.2013; 6. 87-91.
- 229. Singh B, Sharma ML, Gupta DK, Atal CK, Arya RK, Protective effect of Tinospora cordifolia Miers on Carbon tetrachloride induced hepatotoxicity. Indian J Pharmacol 1984, 16 139-142
- 230. Rege N, Dahanukar S & Karandikar S M, Hepatotoxic Effects of Tinospora cordifolia against carbon tetrachloride induced liver damage, Indian Drugs. 1984; 21(12) 544
- 231. Kokate CK & Rambhau D, Antihepatotoxic Activity of Phyllanthus niruri, Tinospora cordifolia and Ricinus communis, Indian Drugs.1993; 30(7) 338.
- 232. Latha U, Rajesh M G & Latha M S, Hepatoprotective Effect of an Ayurvedic Medicine, Indian Drugs. 1999; 36(7), 470
- 233. D. Pandey Protective role of Tinospora cordifolia against lead-induced hepatotoxicity Toxicol. Int. 2010; 17 pp. 12-17
- 234. R. Gupta, V. SharmaA meliorative effects of *Tinospora cordifolia* root extract on histopathological and biochemical changes induced by aflatoxin-b in mice kidney Toxicol. Int. 2011; 18 pp. 94-98
- 235. Ezeonu CS, Egbuna PAC, Ezeanyika LUS, Nkwonta CG, Idoko ND, Antihepatotoxicity studies of crude extract of Zingiber officinale on CCl4 induced toxicity and comparison of the extract's fraction D hepatoprotective capacity, Research Journal of Medical Sciences. 2011; 5(2), 102-107
- 236. Yemitan OK, Izegbu MC.Protective effects of Zingiber officinale (Zingiberaceae) against carbon tetrachloride and acetaminophen-induced hepatotoxicity in rats.Phytother Res. 2006 Nov; 20(11):997-1002.
- 237. Abdullah N, Saat NZM, Hasan HA, Budin SB, Kamaralzaman S, Protective effectof the ethanol extract of Zingiber officinale Roscoe on paracetamol induced hepatotoxicity in rats, Jurnal Sains Kesihatan Malaysia, 2004, 2(2)
- 238. Abdulaziz Bardi D, Halabi MF, Abdullah NA, Rouhollahi E, Hajrezaie M, Abdulla MA. In vivo evaluation of ethanolic extract of Zingiber officinale rhizomes for its protective effect against liver cirrhosis. *Biomed Res Int.* 2013; 2013:918460. doi:10.1155/2013/918460
- 239. Abdelgawad Fahmi,Naglaa Hassanen,Mariam Abdur-Rahman & Engy Shams-Eldin,Phytochemicals, antioxidant activity and hepatoprotective effect of ginger (*Zingiber officinale*) on diethylnitrosamine toxicity in rats, Journal Biomarkers.2019; Volume 24, Issue 5.

- 240. Egwurugwu, J. N. Effects of ginger (Zingiber officinale) on cadmium toxicity, African Journal of Biotechnology. 2007 September; Vol. 6 (18), pp. 2078-2082
- 241. Jharna Bansal, Nitin Kumar, Rishabha Malviya and Pramod Kumar Sharma. Hepatoprotective Models and Various Natural Product Used in Hepatoprotective Agents: a Review, Pharmacognosy Communications. July–Sep 2014; Volume 4, Issue 3
- 242. Yoon, Eric et al. Acetaminophen-Induced Hepatotoxicity: a Comprehensive Update Journal of clinical and translational hepatology. 2016; vol. 4,2. 131-42. doi:10.14218/JCTH.2015.00052
- 243. Torrielli MV, Gabriel L, Dianzani MU. Ethanol-induced hepatotoxicity; experimental observations on the role of lipid peroxidation. J Pathol. 1978 Sep; 126(1):11-25. doi: 10.1002/path.1711260103. PMID: 722405
- 244. Stephanie Mathews, Mingjiang Xu, Hua Wang, Adeline Bertola, Bin Gao, Animals Models of Gastrointestinal and Liver Diseases. Animal models of alcohol-induced liver disease: pathophysiology, translational relevance, and challenges, Am J Physiol Gastrointest Liver Physiol 306: G819–G823, 2014
- 245. MC Wallace , K Hamesch , M Lunova , Y Kim , R Weiskirchen , P Strnad, and SL Friedman, Standard Operating Procedures in Experimental Liver Research: Thioacetamide model in mice and rats, Laboratory Animals 2015, Vol. 49(S1) 21–29
- 246. Yew WW, Leung CC. Antituberculosis drugs and hepatotoxicity. Respirology. 2006 Nov;11(6):699-707. doi: 10.1111/j.1440-1843.2006.00941.x. PMID: 17052297
- 247. Qadrie ZL, Rajkapoor B and Kavimani S: Hepatoprotective Medicinal Herbs and Animal Models for Their Screening A Review. Int J Pharm Sci Res 2015; 6(12): 5006-28.doi: 10.13040/IJPSR.0975-8232.6(12).5006-28
- 248. Ho DW, Lam DK, Chen YB, To J, Ng IO, Fan ST. Galactosamine-induced fulminant liver failure--observation in a porcine model. Asian J Surg. 2002 Jan;25(1):73-9; discussion 80-1. PMID: 17585450
- 249. Tolba R, Kraus T, Liedtke C, Schwarz M, Weiskirchen R. Diethylnitrosamine (DEN)-induced carcinogenic liver injury in mice. Lab Anim. 2015 Apr;49(1 Suppl):59-69. doi: 10.1177/0023677215570086. PMID: 25835739
- 250. Akshatha GM, Raval SK, Arpitha GM, Raval SH, Ghodasara DJ (2018) Immunohistochemical, histopathological study and chemoprotective effect of Solanum nigrum in N-nitrosodiethylamine-induced hepatocellular carcinoma in Wistar rats, Veterinary World, 11(4): 402-409
- 251. Otto Kučera, René Endlicher, Tomáš Roušar, Halka Lotková, Tomáš Garnol, Zdeněk Drahota, Zuzana Červinková, The Effect of *tert*-Butyl Hydroperoxide-Induced Oxidative Stress on Lean and Steatotic Rat Hepatocytes *In Vitro*, *Oxidative Medicine and Cellular Longevity*, vol. 2014, Article ID 752506, 12 pages, 2014. <u>https://doi.org/10.1155/2014/752506</u>
- 252. Pamela D. Moore, Clement G. Yedjou, and Paul B. Tchounwou, Malathion-Induced Oxidative Stress, Cytotoxicity and Genotoxicity in Human Liver Carcinoma (HepG2) Cells, Environ Toxicol. 2010 June ; 25(3): 221–226. doi:10.1002/tox.20492
- 253. Ibrahim, Nabil M et al. Effect of lead acetate toxicity on experimental male albino rat. Asian Pacific journal of tropical biomedicine. 2012; vol. 2, 1. 41-6. doi:10.1016/S2221-1691(11)60187-1
- 254. Bertola A, Mathews S, Ki SH, Wang H, Gao B. Mouse model of chronic and binge ethanol feeding (the NIAAA model). *Nat Protoc*. 2013; 8(3):627-637. doi:10.1038/nprot.2013.032
- 255. Han, Xiao-Dong et al. The involvement of Nrf2 in the protective effects of (-)-Epigallocatechin-3-gallate (EGCG) on NaAsO₂-induced hepatotoxicity. *Oncotarget*. 2017; vol. 8, 39 65302-65312. doi:10.18632/oncotarget.18582

- 256. Lv Y, Hu Q, Shi M, Wang W, Zheng Y, Yang Z, Peng L, Bi D, Zhang A, Hu Y. The role of PSMB5 in sodium arsenite-induced oxidative stress in L-02 cells. Cell Stress Chaperones. 2020 May; 25(3):533-540. doi: 10.1007/s12192-020-01104-1. Epub 2020 Apr 16. PMID: 32301004; PMCID: PMC7192974.
- 257. Cüre, Medine Cumhur et al. Infliximab Modulates Cisplatin-Induced Hepatotoxicity in Rats. *Balkan medical journa*. 2016;*l* vol. 33,5, 504-511. doi:10.5152/balkanmedj.2016.150576
- 258. Vásquez-Garzón, V.R., Ramírez-Cosmes, A., Reyes-Jiménez, E. *et al.* Liver damage in bleomycin-induced pulmonary fibrosis in mice. *Naunyn-Schmiedeberg's Arch Pharmacol.* 2019; 392, 1503–1513. <u>https://doi.org/10.1007/s00210-019-01690-7</u>
- 259. Renugadevi J, Prabu SM. Cadmium-induced hepatotoxicity in rats and the protective effect of naringenin. Exp Toxicol Pathol. 2010 Mar; 62(2):171-81. doi: 10.1016/j.etp.2009.03.010. Epub 2009 May 5. PMID: 19409769
- 260. Kwon J, Kim S, Yoo H, Lee E. Nimesulide-induced hepatotoxicity: A systematic review and meta-analysis. PLoS ONE.2019; 14(1): e0209264. https://doi.org/10.1371/journal.pone.0209264
- 261. Singh, Amrit Pal et al. Mechanisms pertaining to arsenic toxicity. *Toxicology international* vol. 2011;18,2. 87-93. doi:10.4103/0971-6580.84258
- 262. Tan X, Xie G, Sun X, Li Q, Zhong W, Qiao P, Sun X, Jia W, Zhou Z. High fat diet feeding exaggerates perfluorooctanoic acid-induced liver injury in mice via modulating multiple metabolic pathways. PLoS One. 2013 Apr 23; 8(4): e61409. doi: 10.1371/journal.pone.0061409. PMID: 23626681; PMCID: PMC3634078
- 263. Bhatt Narendra, Deshpande Manasi, A Comparative Review On Medicinal Plants Used For The Treatment Of Liver Disorders As In Ayurved, Siddha And Unani [Asu] Systems Of Medicine-Part I- Contextual And Clinical Aspects, International journal of Ayurvedic and Herbal medicine, Vol 11,issue 05, Sep- Oct- 2021
- Achliya GS, Wadodkar SG, Dorle AK. Evaluation of hepatoprotective effect of Amalkadi Ghrita against carbon tetrachloride-induced hepatic damage in rats. J Ethnopharmacol. 2004 Feb;90(2-3):229-32. doi: 10.1016/j.jep.2003.09.037. PMID: 15013185
- 265. Kumar G, Srivastava A, Sharma SK and Gupta YK: Safety evaluation of an Ayurvedic medicine, Arogyavardhini vati on brain, liver and kidney in rats. J Ethnopharmacol, 2012, 140: 151-160.
- 266. Jeena K Jose, Ramadasan Kuttan, Hepatoprotective activity of Emblica officinalis and Chyavanaprash, Journal of Ethnopharmacology. 2000; Volume 72, Issues 1–2, 135-140. https://doi.org/10.1016/S0378-8741 (00)00219-1.
- 267. Deori C, Das S, Bordoloi SK. Study of hepatoprotective activity of Emblica officinalis (AMLA) in Albino rats. J. Evid. Based Med. Healthc. 2017; 4(54), 3298-3301. DOI: 10.18410/jebmh/2017/655
- 268. Dayanand Reddy G, Bernard Mefi M, Bharathy H, Narasimha Kumar GV, Chitikela P Pullaiah, Ganesan R, Sathiya Rajeswaran P. Hepatoprotective Activity of Aruvadha churnam a Traditional Siddha Formulation against Paracetamol Induced Liver Damage in Rats. Research J. Pharm. and Tech. 2018; 11(8): 3380-3384.
- Arunmozhi P.et al, Hepatoprotective Activity of Chara Parpam in Ccl4 Induced Rats, IOSR Journal of Pharmacy and Biological Sciences (IOSR-JPBS). 2013; Volume 5, Issue 6 (Mar. Apr. 2013), 52-58

- 270. Velayudam, Arul Amuthan, Ilavarasan. *Hepatoprotective Activity of Kadukkai Maathirai (A Siddha Polyherbal Formulation) Against Carbon Tetrachloride Induced Liver Damage in Rat.* Research Journal of Pharmaceutical Sciences. 2012; 1 (4). pp. 17-21.
- 271. Kavita Gulati., et al. Hepatoprotective Effects of Dawa-Ul-Kurkum, a Unani Polyherbal Preparation and the Possible Mechanisms in Experimental Model of D-Galactosamine Induced Liver Damage in Rats". EC Pharmacology and Toxicology. 2019; 7.10, 948-960.
- 272. Mehboob Ali, Sagheer Ahmed Khan, Peter S. Chang, Rizwanul Haque, Kanchan Bhatia, Saif Ahmad Habb-e-Asgand, polyherbal Unani formulation, protects liver and antioxidative enzymes against paracetamol induced hepatotoxicity. Pharmaceutical Biology. 2014; 52:4,506-515, DOI: 10.3109/13880209.2013.863949
- 273. Shakya AK, Saxena M, Sharma N, Shrivastava S, Shukla S, .Hepatoprotective Efficacy of Sharbat-e- Deenar against carbon tetrachloride- induced liver damage. 2, 2012, J Environ Pathol Toxicol Oncol, 31: 131- 141
- 274. Girish et. Al : Hepatoprotective Activity of Herbal Formulations In Liver Toxicity, Indian Journal of Experimental Biology. April 2009; Vol. 47, pp. 257-263
- 275. <u>http://www.himalayawellness.com/pro_ducts/pharmaceuticals/liv52</u>.
- 276. Narayanasamy, V. Selvi. Hepatoprotective effect of a polyherbal formulation (Ayush-Liv.04) against ethanol and CCl4 induced liver damage in rats. Ancient Science of Life. 2005; vol. 25, no. 1, pp. 28–33.
- 277. Abul K. Najmi, K. K. Pillai, S. N. Pal, M. Aqil. A. Sayeed. Hepatoprotective and behavioral effects of jigrine in galactosamine-induced hepatopathy in rats, Pharmaceutical Biology. 2010; 48:7, 764-769, DOI: 10.3109/13880200903300188
- 278. Mahim Zameer, Abdur Rauf, Iqbal Ahmad Qasm. Hepatoprotective Activity of a Unani Polyherbal Formulation Kabideen in Paracetamol Induced Liver Toxicity in Rats. Hippocratic Journal of Unani Medicine. October - December 2014; Vol. 9 No. 4, Pages 41-50
- 279. Debendranath Dey, Sunetra Chaskar, Narendra Bhatt, Deepa Chitre, "Hepatoprotective Activity of BV-7310, a Proprietary Herbal Formulation of *Phyllanthus niruri*, *Tephrosia purpurea*, *Boerhavia diffusa*, and *Andrographis paniculata*, in Alcohol-Induced HepG2 Cells and Alcohol plus a Haloalkane, CCl4, Induced Liver Damage in Rats", *Evidence-Based Complementary and Alternative Medicine*, vol. 2020, Article ID 6428906, 9 pages. https://doi.org/10.1155/2020/6428906
- 280. Kohli K.R. Tathed P.S., Bhatt N.S, Effect of An Ayurvedic Drug L 2002@ in Viral Hepatitis, The Indian Practitioner, Vol XLVIII, No 7, pp 635-642, July 1995.
- 281. Deshpande P.J., Singh R , Bhatt N.S, Clinical Effect of An Ayurvedic Drug L 2002 @ In Hepato-Biliary Disorders, Journal of National Integrated Medical Association, Vol. XXXVI, No.12, pp.5-11, December 1994
- 282. N.S. Bhatt, Banavalikar S, Ghoda M, Evaluation of Hepafyte (A Herral Preparation) for Alcoholic Liver Disease, <u>http://www.drnarendrabhatt.in/researchpapers.html</u>
- 283. Tiwari P, Kumar B, Kaur M, Kaur G, Kaur H. Phytochemical screening and extraction: A review. *Int Pharm Sci.* 2011; 1:98–106.
- 284. Pandey A, Tripathi S. Concept of standardization, extraction, and pre-phytochemical screening strategies for herbal drug. *J Pharmacogn Phytochem.* 2014; 2:115–9.
- 285. S. Shailajan, M. Joshi, B. Tiwari, **Hepatoprotective activity of** *Parmelia perlata* (**Huds.**) Ach. against CCl4 induced liver toxicity in Albino Wistar rats, J Appl Pharm Sci, (2014), 4. pp. 70-74. 10.7324/JAPS.2014.40212

 Kumar D, Patil PA, Roy S, Kholkute SD, Hegde HV, Nair V. Comparative toxicity profiles of Plumbago zeylanica L. root petroleum ether, acetone and hydroalcoholic extracts in Wistar rats. Ayu. 2015 Jul-Sep;36(3):329-34. doi: 10.4103/0974-8520.182750. PMID: 27313422; PMCID: PMC4895762.